



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C12N 15/53, 15/11, 15/82, 1/21, 5/10, 9/02, C12Q 1/68, A01H 5/00		A1	(11) International Publication Number: WO 98/56922 (43) International Publication Date: 17 December 1998 (17.12.98)																																				
<p>(21) International Application Number: PCT/US98/12074</p> <p>(22) International Filing Date: 11 June 1998 (11.06.98)</p> <p>(30) Priority Data: 08/872,302 11 June 1997 (11.06.97) US</p> <p>(63) Related by Continuation (CON) or Continuation-in-Part (CIP) to Earlier Application US 08/872,302 (CIP) Filed on 11 June 1997 (11.06.97)</p> <p>(71) Applicant (for all designated States except US): E.I. DU PONT DE NEMOURS AND COMPANY [US/US]; 1007 Market Street, Wilmington, DE 19898 (US).</p> <p>(72) Inventor; and (75) Inventor/Applicant (for US only): HITZ, William, D. [US/US]; 404 Hillside Road, Wilmington, DE 19807 (US).</p> <p>(74) Agent: MAJARIAN, William, R.; E.I. du Pont de Nemours and Company, Legal Patent Records Center, 1007 Market Street, Wilmington, DE 19898 (US).</p>		<p>(81) Designated States: AL, AM, AU, AZ, BA, BB, BG, BR, BY, CA, CN, CU, CZ, EE, GE, GW, HU, ID, IL, IS, JP, KG, KP, KR, KZ, LC, LK, LR, LT, LV, MD, MG, MK, MN, MX, NO, NZ, PL, RO, RU, SG, SI, SK, SL, TJ, TM, TR, TT, UA, US, UZ, VN, YU, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).</p> <p>Published With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments. With an indication in relation to deposited biological material furnished under Rule 13bis separately from the description.</p>																																					
<p>(54) Title: FATTY ACID MODIFYING ENZYMES FROM DEVELOPING SEEDS OF VERNONIA GALAMENENSIS</p> <p>(57) Abstract</p> <p>This invention relates to isolated nucleic acid fragments encoding all or a substantial portion of <i>Vernonia galamenensis</i> fatty acid modifying enzymes. The invention also relates to the construction of chimeric genes encoding all or a portion of <i>Vernonia galamenensis</i> fatty acid modifying enzymes, in sense or antisense orientation, wherein expression of the chimeric gene results in production of altered levels of a <i>Vernonia galamenensis</i> fatty acid modifying enzymes in transformed host cells.</p> <table border="1"> <caption>Estimated data for Figure 1</caption> <thead> <tr> <th>SEEDS CLASS</th> <th>18:1 12, 13 epoxy fatty acid (% of total fatty acids)</th> <th>Frequency</th> </tr> </thead> <tbody> <tr><td>0-1</td><td>0.5</td><td>32</td></tr> <tr><td>1-2</td><td>1.0</td><td>2</td></tr> <tr><td>2-3</td><td>1.5</td><td>1</td></tr> <tr><td>3-4</td><td>2.0</td><td>1</td></tr> <tr><td>4-5</td><td>3.5</td><td>10</td></tr> <tr><td>5-6</td><td>4.5</td><td>14</td></tr> <tr><td>6-7</td><td>5.5</td><td>13</td></tr> <tr><td>7-8</td><td>7.5</td><td>9</td></tr> <tr><td>8-9</td><td>8.5</td><td>2</td></tr> <tr><td>9-10</td><td>9.5</td><td>1</td></tr> <tr><td>10-11</td><td>10.5</td><td>1</td></tr> </tbody> </table>				SEEDS CLASS	18:1 12, 13 epoxy fatty acid (% of total fatty acids)	Frequency	0-1	0.5	32	1-2	1.0	2	2-3	1.5	1	3-4	2.0	1	4-5	3.5	10	5-6	4.5	14	6-7	5.5	13	7-8	7.5	9	8-9	8.5	2	9-10	9.5	1	10-11	10.5	1
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TITLEFATTY ACID MODIFYING ENZYMES FROM DEVELOPING
SEEDS OF *VERNONIA GALAMENENSIS*FIELD OF THE INVENTION

5 This invention is in the field of plant molecular biology. More specifically, this invention pertains to nucleic acid fragments encoding enzymes involved in fatty acid biosynthesis and modification in plants and seeds.

BACKGROUND OF THE INVENTION

10 Fatty acids bearing chemical modifications in addition to the common double bonds are found in the storage lipids of many oilseeds (Harwood, J. L. (1980) *In* The Biochemistry of Plants, T.S. Moore Jr., *ed.* CRC Press, New York, pp 91-116). Some of these 15 modifications functionalize the fatty acid to produce products that are useful in industrial applications; this is opposed to the more common usage of plant-derived lipids as foods. Examples are the use of the hydroxylated fatty acid ricinoleic acid in lubricants, and the short- or medium-carbon chain length fatty acids from palm oil in detergents. In some cases, fatty acid composition of the storage lipids of oilseeds produced in temperate climates can be modified by the addition of genes from exotic sources so that large amounts of unique fatty acids are produced (Ohlrogge, J. B. (1994) *Plant Physiol.* 104, 821-826).

20 Epoxidation is among the known modifications to storage lipid fatty acids. The 18-carbon fatty acid 9,10-ene-12,13-epoxide comprises as much as 60% of the total seed fatty acid in species such as *Vernonia galamenensis* and *Euphorbia lagascea* (Bafor, M. et al. (1993) *Arch. Biochem Biophys* 303:145-151). Fatty acids carrying the epoxide modification may find use as plasticizers, in crosslinking coatings applications, and in setting printing inks.

25 Attempts to delineate the biosynthetic pathway of cis-12-epoxyoctadeca-cis -9-enoate indicate that the catalytic activity responsible for the introduction of the epoxide moiety is in the microsomal membrane fraction, most likely the endoplasmic reticulum (Bafor et al. *supra*). While the above study also suggests that the catalytic activity responsible is an enzyme in the cytochrome P450 mono-oxygenase class, enzymes with amino acid sequences 30 related to the endoplasmic reticulum-localized fatty acid desaturases have been isolated from tissues that produce hydroxylated fatty acids (World Patent Publication No. WO94/11516). These sequences have been shown to be active in adding the hydroxyl group to esterified fatty acids (Broun, P. and Somerville, C. (1997) *Plant Physiol* 113: 933-942). It is therefore possible that fatty acid epoxidizing enzymes may be related in sequence to the class of 35 membrane bound enzymes responsible for fatty acid desaturation and fatty acid hydroxylation.

Thus while candidate enzyme classes have been suggested, no gene sequences from those candidate classes and from tissues that are known to produce epoxidized fatty acids have been isolated.

SUMMARY OF THE INVENTION

5 The instant invention relates to isolated nucleic acid fragments encoding plant enzymes involved in fatty acid biosynthesis and modification, the enzymes having sequence homology to membrane-bound fatty acid desaturases. Specifically, this invention concerns isolated nucleic acid fragments encoding a fatty acid epoxidizing enzyme and a fatty acid desaturase enzyme, each of which are normally expressed in developing seeds of *Vernonia galamenensis*. In addition, this invention relates to nucleic acid fragments that are 10 complementary to nucleic acid fragments encoding the *Vernonia galamenensis* fatty acid epoxidizing and desaturase enzymes.

In another embodiment, the instant invention relates chimeric genes that comprise 15 nucleic acid fragments encoding *Vernonia galamenensis* fatty acid epoxidizing or desaturase enzymes operably linked to suitable regulatory sequences, wherein expression of the chimeric genes results in production of levels of the encoded protein in transformed host cells. For example, disclosed herein is a chimeric gene wherein a nucleic acid fragment 20 encoding a *Vernonia galamenensis* fatty acid epoxidizing enzyme or a *Vernonia galamenensis* fatty acid desaturase enzyme is operably linked to one or more regulatory sequences suitable for directing expression of the *Vernonia galamenensis* fatty acid epoxidizing or desaturase enzymes in microbial cells.

In a further embodiment, the instant invention concerns a transformed host cell comprising in its genome a chimeric gene comprising a nucleic acid fragment encoding a *Vernonia galamenensis* fatty acid epoxidizing or desaturase enzyme operably linked to 25 suitable regulatory sequences. Expression of the chimeric gene results in production of altered levels of protein encoded by the operably linked nucleic acid fragment in the transformed host cell. The transformed host cell can be of eukaryotic or prokaryotic origin, and include cells derived from higher plants and microorganisms. The invention also includes transformed plants that arise from transformed host cells of higher plants, and from 30 seeds derived from such transformed plants.

A further embodiment of the instant invention is a soybean seed that accumulates vernolic acid.

An additional embodiment of the instant invention concerns a method of altering the level of expression of a *Vernonia galamenensis* fatty acid epoxidizing enzyme or a *Vernonia galamenensis* fatty acid desaturase enzyme in a transformed host cell comprising: 35 a) transforming a host cell with a chimeric gene comprising a nucleic acid fragment encoding a *Vernonia galamenensis* fatty acid epoxidizing enzyme or a *Vernonia galamenensis* fatty acid desaturase enzyme; and b) growing the transformed host cell under conditions that are

suitable for expression of the chimeric gene wherein expression of the chimeric gene results in production of altered levels of protein encoded by the operably linked nucleic acid fragment in the transformed host cell.

Yet another embodiment of the instant invention concerns a method for producing 5 epoxidized fatty acids in the seeds of plants comprising the following steps: (a) transforming a plant cell with a chimeric gene encoding all or a portion of the *Vernonia galamenensis* fatty acid epoxidizing enzyme operably linked in sense orientation to suitable regulatory sequences; (b) growing a fertile mature plant from the transformed plant cell of step (a) under conditions suitable to obtain seeds; and (c) selecting from the progeny seed of step (b) 10 those seeds containing epoxidized fatty acids.

Another embodiment of the instant invention is a method for producing *Vernonia galamenensis* fatty acid epoxidizing enzyme or *Vernonia galamenensis* fatty acid desaturase enzyme comprising the following steps: (a) transforming a microbial host cell with a chimeric gene wherein a nucleic acid fragment encoding a *Vernonia galamenensis* fatty acid 15 epoxidizing enzyme or a *Vernonia galamenensis* fatty acid desaturase enzyme is operably linked to regulatory sequences suitable for directing expression in microbial cells; and (b) growing the transformed microbial cells obtained from step (a) under conditions that result in expression of the *Vernonia galamenensis* fatty acid epoxidizing or desaturase enzyme proteins.

20 An additional embodiment of the instant invention concerns a method for obtaining a nucleic acid fragment encoding all or substantially all of an amino acid sequence encoding *Vernonia galamenensis* fatty acid epoxidizing or desaturase enzymes.

BRIEF DESCRIPTION OF THE
DRAWINGS AND SEQUENCE DESCRIPTIONS

25 The invention can be more fully understood from the following detailed description and the accompanying drawing and sequence descriptions which form a part of this application.

Figure 1 shows a comparison of the amino acid sequences of the instant *Vernonia galamenensis* fatty acid desaturase (vs1.05h08), the instant *Vernonia galamenensis* fatty acid 30 epoxidase (vs1.02c07), a soybean fatty acid desaturase (soy) and a castor bean fatty acid hydroxylase (castor). The eight conserved histidine residues found in all membrane bound fatty acid modifying enzymes of this class are boxed and stippled. An arginine residue found in the fatty acid hydroxylase from castor bean and in the instant *Vernonia galamenensis* fatty acid epoxidase but not in the fatty acid desaturating enzymes of this class 35 is also boxed and stippled. Other residues in the *Vernonia* fatty acid epoxidase sequence that are unique to that sequence but conserved in most or all of the other sequences are boxed. Not all amino acid changes are marked: only those in which the change in the epoxidase sequence occurs in a highly conserved region and in which the amino acid present in the

epoxidase is different in physical character from the conserved residues in the other sequences are marked.

Figure 2 is a restriction map of plasmid pRVF7 used for soybean transformations.

5 Figure 3 is a histogram of the results of single seed analyses of the accumulation of vernolic acid in seeds from plants transformed with pRVF7.

The following sequence descriptions and sequence listings attached hereto comply with the rules governing nucleotide and/or amino acid sequence disclosures in patent applications as set forth in 37 C.F.R. §1.821-1.825.

10 SEQ ID NO:1 is the nucleotide sequence comprising the cDNA insert in clone vs1.05h08 encoding a *Vernonia galamenensis* fatty acid desaturase enzyme.

SEQ ID NO:2 is the deduced amino acid sequence of a *Vernonia galamenensis* fatty acid desaturase enzyme derived from the nucleotide sequence of SEQ ID NO:1.

15 SEQ ID NO:3 is the nucleotide sequence comprising the cDNA insert in clone vs1.02c07 encoding a *Vernonia galamenensis* fatty acid epoxidizing enzyme.

SEQ ID NO:4 is the deduced amino acid sequence of a *Vernonia galamenensis* fatty acid epoxidizing enzyme derived from the nucleotide sequence of SEQ ID NO:3.

SEQ ID NO:5 is the amino acid sequence encoding the soybean (*Glycine max*) fatty acid desaturase enzyme depicted in Figure 1 and having GenBank Accession No. L43920.

20 SEQ ID NO:6 is the amino acid sequence encoding the castor bean (*Ricinus communis*) fatty acid hydroxylase enzyme depicted in Figure 1 and having GenBank Accession No. U22378.

SEQ ID NO:7 shows the nucleotide sequence of the PCR primer used as the 5' end primer in PCR reactions for amplification of the coding region of vs1.02c07.

25 SEQ ID NO:8 shows the nucleotide sequence of the PCR primer used as the 3' end primer in PCR reactions for amplification of the coding region of vs1.02c07.

SEQ ID NO:9 shows the nucleotide sequence of the PCR primer used as the 5' end primer in PCR reactions for amplification of the coding region of vs1.05h08.

SEQ ID NO:10 shows the nucleotide sequence of the PCR primer used as the 3' end primer in PCR reactions for amplification of the coding region of vs1.05h08.

30 The Sequence Descriptions contain the one letter code for nucleotide sequence characters and the three letter codes for amino acids as defined in conformity with the IUPAC-IYUB standards described in *Nucleic Acids Research* 13:3021-3030 (1985) and in the *Biochemical Journal* 219 (No. 2):345-373 (1984) which are herein incorporated by reference. The symbols and format used for nucleotide and amino acid sequence data comply with the rules set forth in 37 C.F.R. §1.822.

BIOLOGICAL DEPOSIT

The following soybean seed has been deposited with the American Type Culture Collection (ATCC), 10801 University Boulevard, Manassas, VA 20110-2209, and bears the following designation, accession number and date of deposit.

5

Designation	Accession Number	Date of Deposit
798	ATCC XXXXX	June __, 1998

DETAILED DESCRIPTION OF THE INVENTION

This invention relates to isolated nucleic acid fragments encoding all or a substantial portion of fatty acid epoxidizing and desaturase enzymes normally expressed in developing seeds of *Vernonia galamenensis* that are similar in sequence to other plant, membrane-bound fatty acid desaturases. The invention also relates to the construction of a chimeric gene comprising an nucleic acid fragment encoding all or a portion of the *Vernonia galamenensis* fatty acid epoxidizing or desaturase enzyme, operably linked in sense or antisense orientation to suitable regulatory sequences, wherein expression of the chimeric gene results in 10 production of altered levels of the desired enzyme in a transformed host cell. The invention also relates to methods of using isolated nucleic acid fragments encoding all or a substantial portion of the fatty acid epoxidizing and desaturase enzymes normally expressed in 15 developing seeds of *Vernonia galamenensis*.

In the context of this disclosure, a number of terms shall be utilized. As used herein, 20 an "isolated nucleic acid fragment" is a polymer of RNA or DNA that is single- or double-stranded, optionally containing synthetic, non-natural or altered nucleotide bases. An isolated nucleic acid fragment in the form of a polymer of DNA may be comprised of one or more segments of cDNA, genomic DNA or synthetic DNA.

The term "*Vernonia galamenensis* fatty acid modifying enzyme" refers collectively to 25 the *Vernonia galamenensis* fatty acid epoxidizing enzyme and the *Vernonia galamenensis* fatty acid desaturase enzyme disclosed in the instant specification.

As used herein, "substantially similar" refers to nucleic acid fragments wherein 30 changes in one or more nucleotides results in substitution of one or more amino acids, but do not affect the functional properties of the protein encoded by the DNA sequence. "Substantially similar" also refers to nucleic acid fragments wherein changes in one or more nucleotides does not affect the ability of the nucleic acid fragment to mediate alteration of 35 gene expression by antisense or co-suppression technology. "Substantially similar" also refers to modifications of the nucleic acid fragments of the instant invention such as deletion or insertion of one or more nucleotides that do not substantially affect the functional properties of the resulting transcript vis-à-vis the ability to mediate alteration of gene expression by antisense or co-suppression technology or alteration of the functional

properties of the resulting protein molecule. It is therefore understood that the invention encompasses more than the specific exemplary sequences.

For example, it is well known in the art that antisense suppression and co-suppression of gene expression may be accomplished using nucleic acid fragments representing less than the entire coding region of a gene, and by nucleic acid fragments that do not share 100% identity with the gene to be suppressed. Moreover, alterations in a gene which result in the production of a chemically equivalent amino acid at a given site, but do not effect the functional properties of the encoded protein, are well known in the art. Thus, a codon for the amino acid alanine, a hydrophobic amino acid, may be substituted by a codon encoding another less hydrophobic residue, such as glycine, or a more hydrophobic residue, such as valine, leucine, or isoleucine. Similarly, changes which result in substitution of one negatively charged residue for another, such as aspartic acid for glutamic acid, or one positively charged residue for another, such as lysine for arginine, can also be expected to produce a functionally equivalent product. Nucleotide changes which result in alteration of the N-terminal and C-terminal portions of the protein molecule would also not be expected to alter the activity of the protein. Each of the proposed modifications is well within the routine skill in the art, as is determination of retention of biological activity of the encoded products. Moreover, the skilled artisan recognizes that substantially similar sequences encompassed by this invention are also defined by their ability to hybridize, under stringent conditions (0.1X SSC, 0.1% SDS, 65°C), with the sequences exemplified herein. Preferred substantially similar nucleic acid fragments of the instant invention are those nucleic acid fragments whose DNA sequences are 80% identical to the DNA sequence of the nucleic acid fragments reported herein. More preferred nucleic acid fragments are 90% identical to the DNA sequence of the nucleic acid fragments reported herein. Most preferred are nucleic acid fragments that are 95% identical to the DNA sequence of the nucleic acid fragments reported herein.

A "substantial portion" of an amino acid or nucleotide sequence comprises enough of the amino acid sequence of a polypeptide or the nucleotide sequence of a gene to afford putative identification of that polypeptide or gene, either by manual evaluation of the sequence by one skilled in the art, or by computer-automated sequence comparison and identification using algorithms such as BLAST (Basic Local Alignment Search Tool; Altschul, S. F., et al., (1993) *J. Mol. Biol.* 215:403-410; see also www.ncbi.nlm.nih.gov/BLAST/). In general, a sequence of ten or more contiguous amino acids or thirty or more nucleotides is necessary in order to putatively identify a polypeptide or nucleic acid sequence as homologous to a known protein or gene. Moreover, with respect to nucleotide sequences, gene specific oligonucleotide probes comprising 20-30 contiguous nucleotides may be used in sequence-dependent methods of gene identification (e.g., Southern hybridization) and isolation (e.g., *in situ* hybridization of bacterial colonies or

bacteriophage plaques). In addition, short oligonucleotides of 12-15 bases may be used as amplification primers in PCR in order to obtain a particular nucleic acid fragment comprising the primers. Accordingly, a "substantial portion" of an amino acid or nucleotide sequence comprises enough of the sequence to afford specific identification and/or isolation

5 of a nucleic acid fragment comprising the sequence. The instant specification teaches partial or complete amino acid and nucleotide sequences encoding one or more particular plant proteins. The skilled artisan, having the benefit of the sequences as reported herein, may now use all or a substantial portion of the disclosed sequences for purposes known to those skilled in this art. Accordingly, the instant invention comprises the complete sequences as

10 reported in the accompanying Sequence Listing, as well as substantial portions of those sequences as defined above.

"Codon degeneracy" refers to divergence in the genetic code permitting variation of the nucleotide sequence without effecting the amino acid sequence of an encoded polypeptide. Accordingly, the instant invention relates to any nucleic acid fragment that encodes all or a substantial portion of the amino acid sequence encoding the *Vernonia galamenensis* fatty acid desaturase enzyme as set forth in SEQ ID NO:2, and to any nucleic acid fragment that encodes all or a substantial portion of the amino acid sequence encoding the *Vernonia galamenensis* fatty acid epoxidizing enzyme as set forth in SEQ ID NO:4. The skilled artisan is well aware of the "codon-bias" exhibited by a specific host cell in usage of nucleotide codons to specify a given amino acid. Therefore, when synthesizing a gene for improved expression in a host cell, it is desirable to design the gene such that its frequency of codon usage approaches the frequency of preferred codon usage of the host cell.

"Synthetic genes" can be assembled from oligonucleotide building blocks that are chemically synthesized using procedures known to those skilled in the art. These building blocks are ligated and annealed to form gene segments which are then enzymatically assembled to construct the entire gene. "Chemically synthesized", as related to a sequence of DNA, means that the component nucleotides were assembled *in vitro*. Manual chemical synthesis of DNA may be accomplished using well established procedures, or automated chemical synthesis can be performed using one of a number of commercially available machines. Accordingly, the genes can be tailored for optimal gene expression based on optimization of nucleotide sequence to reflect the codon bias of the host cell. The skilled artisan appreciates the likelihood of successful gene expression if codon usage is biased towards those codons favored by the host. Determination of preferred codons can be based on a survey of genes derived from the host cell where sequence information is available.

35 "Gene" refers to a nucleic acid fragment that expresses a specific protein, including regulatory sequences preceding (5' non-coding sequences) and following (3' non-coding sequences) the coding sequence. "Native gene" refers to a gene as found in nature with its own regulatory sequences. "Chimeric gene" refers any gene that is not a native gene,

comprising regulatory and coding sequences that are not found together in nature.

Accordingly, a chimeric gene may comprise regulatory sequences and coding sequences that are derived from different sources, or regulatory sequences and coding sequences derived from the same source, but arranged in a manner different than that found in nature.

5 "Endogenous gene" refers to a native gene in its natural location in the genome of an organism. A "foreign" gene refers to a gene not normally found in the host organism, but that is introduced into the host organism by gene transfer. Foreign genes can comprise native genes inserted into a non-native organism, or chimeric genes. A "transgene" is a gene that has been introduced into the genome by a transformation procedure.

10 "Coding sequence" refers to a DNA sequence that codes for a specific amino acid sequence. "Regulatory sequences" refer to nucleotide sequences located upstream (5' non-coding sequences), within, or downstream (3' non-coding sequences) of a coding sequence, and which influence the transcription, RNA processing or stability, or translation of the associated coding sequence. Regulatory sequences may include promoters, translation 15 leader sequences, introns, and polyadenylation recognition sequences.

"Promoter" refers to a DNA sequence capable of controlling the expression of a coding sequence or functional RNA. In general, a coding sequence is located 3' to a promoter sequence. The promoter sequence consists of proximal and more distal upstream elements, the latter elements often referred to as enhancers. Accordingly, an "enhancer" is a 20 DNA sequence which can stimulate promoter activity and may be an innate element of the promoter or a heterologous element inserted to enhance the level or tissue-specificity of a promoter. Promoters may be derived in their entirety from a native gene, or be composed of different elements derived from different promoters found in nature, or even comprise synthetic DNA segments. It is understood by those skilled in the art that different promoters 25 may direct the expression of a gene in different tissues or cell types, or at different stages of development, or in response to different environmental conditions. Promoters which cause a gene to be expressed in most cell types at most times are commonly referred to as "constitutive promoters". New promoters of various types useful in plant cells are constantly being discovered; numerous examples may be found in the compilation by 30 Okamuro and Goldberg, (1989) *Biochemistry of Plants* 15:1-82. It is further recognized that since in most cases the exact boundaries of regulatory sequences have not been completely defined, DNA fragments of different lengths may have identical promoter activity.

The "translation leader sequence" refers to a DNA sequence located between the promoter sequence of a gene and the coding sequence. The translation leader sequence is 35 present in the fully processed mRNA upstream of the translation start sequence. The translation leader sequence may affect processing of the primary transcript to mRNA, mRNA stability or translation efficiency. Examples of translation leader sequences have been described (Turner, R. and Foster, G.D. (1995) *Molecular Biotechnology* 3:225).

The "3' non-coding sequences" refer to DNA sequences located downstream of a coding sequence and include polyadenylation recognition sequences and other sequences encoding regulatory signals capable of affecting mRNA processing or gene expression. The polyadenylation signal is usually characterized by affecting the addition of polyadenylic acid tracts to the 3' end of the mRNA precursor. The use of different 3' non-coding sequences is exemplified by Ingelbrecht et al., (1989) *Plant Cell* 1:671-680.

"RNA transcript" refers to the product resulting from RNA polymerase-catalyzed transcription of a DNA sequence. When the RNA transcript is a perfect complementary copy of the DNA sequence, it is referred to as the primary transcript; or it may be a RNA sequence derived from posttranscriptional processing of the primary transcript and is referred to as the mature RNA. "Messenger RNA" (mRNA) refers to the RNA that is without introns and that can be translated into protein by the cell. "cDNA" refers to a double-stranded DNA that is complementary to and derived from mRNA. "Sense" RNA refers to RNA transcript that includes the mRNA and so can be translated into protein by the cell. "Antisense RNA" refers to a RNA transcript that is complementary to all or part of a target primary transcript or mRNA and that blocks the expression of a target gene (U.S. Pat. No. 5,107,065). The complementarity of an antisense RNA may be with any part of the specific gene transcript, i.e., at the 5' non-coding sequence, 3' non-coding sequence, introns, or the coding sequence. "Functional RNA" refers to antisense RNA, ribozyme RNA, or other RNA that is not translated yet has an effect on cellular processes.

The term "operably linked" refers to the association of nucleic acid sequences on a single nucleic acid fragment so that the function of one is affected by the other. For example, a promoter is operably linked with a coding sequence when it is capable of affecting the expression of that coding sequence (i.e., that the coding sequence is under the transcriptional control of the promoter). Coding sequences can be operably linked to regulatory sequences in sense or antisense orientation.

The term "expression", as used herein, refers to the transcription and stable accumulation of sense (mRNA) or antisense RNA derived from the nucleic acid fragment of the invention. Expression may also refer to translation of mRNA into a polypeptide. "Antisense inhibition" refers to the production of antisense RNA transcripts capable of suppressing the expression of the target protein. "Overexpression" refers to the production of a gene product in transgenic organisms that exceeds levels of production in normal or non-transformed organisms. "Co-suppression" refers to the production of sense RNA transcripts capable of suppressing the expression of identical or substantially similar foreign or endogenous genes (U.S. Pat. No. 5,231,020).

"Altered levels" refers to the production of gene product(s) in transgenic organisms in amounts or proportions that differ from that of normal or non-transformed organisms.

“Mature” protein refers to a post-translationally processed polypeptide; i.e., one from which any pre- or propeptides present in the primary translation product have been removed. “Precursor” protein refers to the primary product of translation of mRNA; i.e., with pre- and propeptides still present. Pre- and propeptides may be but are not limited to intracellular localization signals.

5 A “chloroplast transit peptide” is an amino acid sequence which is translated in conjunction with a protein and directs the protein to the chloroplast or other plastid types present in the cell in which the protein is made. “Chloroplast transit sequence” refers to a nucleotide sequence that encodes a chloroplast transit peptide. A “signal peptide” is an 10 amino acid sequence which is translated in conjunction with a protein and directs the protein to the secretory system (Chrispeels, J.J., (1991) *Ann. Rev. Plant Phys. Plant Mol. Biol.* 42:21-53). If the protein is to be directed to a vacuole, a vacuolar targeting signal (*supra*) can further be added, or if to the endoplasmic reticulum, an endoplasmic reticulum retention signal (*supra*) may be added. If the protein is to be directed to the nucleus, any signal 15 peptide present should be removed and instead a nuclear localization signal included (Raikhel (1992) *Plant Phys.* 100:1627-1632).

“Transformation” refers to the transfer of a nucleic acid fragment into the genome of a host organism, resulting in genetically stable inheritance. Host organisms containing the transformed nucleic acid fragments are referred to as “transgenic” organisms. Examples of 20 methods of plant transformation include Agrobacterium-mediated transformation (De Blaere et al. (1987) *Meth. Enzymol.* 143:277) and particle-accelerated or “gene gun” transformation technology (Klein et al. (1987) *Nature (London)* 327:70-73; U.S. Pat. No. 4,945,050).

25 Standard recombinant DNA and molecular cloning techniques used herein are well known in the art and are described more fully in Sambrook, J., Fritsch, E.F. and Maniatis, T. *Molecular Cloning: A Laboratory Manual*; Cold Spring Harbor Laboratory Press: Cold Spring Harbor, 1989 (hereinafter “Maniatis”).

This invention relates to plant cDNAs with homology to fatty acid desaturase enzymes 30 from other plant species. Several cDNA clones encoding *Vernonia galamenensis* fatty acid modifying enzymes have been isolated and identified by comparison of random plant cDNA sequences to the GenBank database using the BLAST algorithms well known to those skilled in the art. The nucleotide sequence encoding the *Vernonia galamenensis* fatty acid desaturase enzyme is provided in SEQ ID NO:1, and the deduced amino acid sequence is provided in SEQ ID NO:2. The nucleotide sequence encoding the *Vernonia galamenensis* fatty acid epoxidizing enzyme is provided in SEQ ID NO:3, and the deduced amino acid 35 sequence is provided in SEQ ID NO:4. Fatty acid desaturase and epoxidizing enzymes genes from other plants can now be identified by comparison of random cDNA sequences to the *Vernonia galamenensis* sequences provided herein.

The amino acid sequences encoded by the cDNA clones disclosed herein are compared in Figure 1 to the fatty acid desaturase from soybean which inserts the second double bond between carbon atoms 12 and 13 into mono-unsaturated fatty acid, oleic acid, to produce linoleic acid. The sequence of a similar enzyme from castor bean which functions to

5 hydroxylate the number 12 carbon atom of oleic acid to produce ricinoleic acid is also shown for comparison. While both *Vernonia galamenensis* sequences possess the highly conserved amino acid residues that are common to this class of enzyme, one of the clones, vs1.05h08, demonstrates greater similarity to the soybean sequence than vs1.02c07. The amino acid sequence encoded by cDNA clone vs1.05h08 is 70.2% similar to the soybean sequence,

10 while the sequence encoded by cDNA clone vs1.02c07 is only 53.8% similar. As well, the two *Vernonia galamenensis* sequences show only 57.7% similarity to each other as opposed to the much greater sequence similarity shown between species by vs1.05h08.

In *Vernonia galamenensis*, as with other species that produce unusual fatty acids, production of these unusual fatty acids is limited to seed storage tissue. Moreover, the

15 unusual fatty acids are generally not found in other parts of the plant. No signal was detected following Northern analysis of mRNA isolated from leaves of *Vernonia galamenensis* when vs1.02c07 was used as a probe, while the message was very abundant in mRNA isolated from developing seeds.

The sequence relationship of vs1.02c07 to other, known fatty acid desaturases, along

20 with its tissue-specific expression pattern in a tissue that is actively producing the epoxidized fatty acid vernoleate, make it very likely that the enzyme encoded by cDNA clone vs1.02c07 is a fatty acid epoxidase. In contrast, sequence comparisons indicate that cDNA clone vs1.05h08 encodes a fatty acid desaturase enzyme.

The nucleic acid fragments of the instant invention may be used to isolate cDNAs and

25 genes encoding homologous fatty acid modifying enzymes from the same or other plant species. Isolation of homologous genes using sequence-dependent protocols is well known in the art. Examples of sequence-dependent protocols include, but are not limited to, methods of nucleic acid hybridization, and methods of DNA and RNA amplification as exemplified by various uses of nucleic acid amplification technologies (e.g., polymerase

30 chain reaction, ligase chain reaction).

For example, genes encoding homologous fatty acid modifying enzymes, either as

cDNAs or genomic DNAs, could be isolated directly by using all or a portion of the instant

nucleic acid fragments as DNA hybridization probes to screen libraries from any desired

plant employing methodology well known to those skilled in the art. Specific

35 oligonucleotide probes based upon the instant nucleic acid sequences can be designed and synthesized by methods known in the art (Maniatis). Moreover, the entire sequences can be used directly to synthesize DNA probes by methods known to the skilled artisan such as random primers DNA labeling, nick translation, or end-labeling techniques, or RNA probes

using available *in vitro* transcription systems. In addition, specific primers can be designed and used to amplify a part of or full-length of the instant sequences. The resulting amplification products can be labeled directly during amplification reactions or labeled after amplification reactions, and used as probes to isolate full length cDNA or genomic fragments under conditions of appropriate stringency.

5 In addition, two short segments of the instant nucleic acid fragments may be used in polymerase chain reaction protocols to amplify longer nucleic acid fragments encoding homologous genes from DNA or RNA. The polymerase chain reaction may also be performed on a library of cloned nucleic acid fragments wherein the sequence of one primer
10 is derived from the instant nucleic acid fragments, and the sequence of the other primer takes advantage of the presence of the polyadenylic acid tracts to the 3' end of the mRNA precursor encoding plant genes. Alternatively, the second primer sequence may be based upon sequences derived from the cloning vector. For example, the skilled artisan can follow the RACE protocol (Frohman et al., (1988) *PNAS USA* 85:8998) to generate cDNAs by
15 using PCR to amplify copies of the region between a single point in the transcript and the 3' or 5' end. Primers oriented in the 3' and 5' directions can be designed from the instant sequences. Using commercially available 3' RACE or 5' RACE systems (BRL), specific 3' or 5' cDNA fragments can be isolated (Ohara et al., (1989) *PNAS USA* 86:5673; Loh et al., (1989) *Science* 243:217). Products generated by the 3' and 5' RACE procedures can be
20 combined to generate full-length cDNAs (Frohman, M.A. and Martin, G.R., (1989) *Techniques* 1:165).

25 Availability of the instant nucleotide and deduced amino acid sequences facilitates immunological screening cDNA expression libraries. Synthetic peptides representing portions of the instant amino acid sequences may be synthesized. These peptides can be used to immunize animals to produce polyclonal or monoclonal antibodies with specificity for peptides or proteins comprising the amino acid sequences. These antibodies can be then be used to screen cDNA expression libraries to isolate full-length cDNA clones of interest (Lerner, R.A. (1984) *Adv. Immunol.* 36:1; Maniatis).

30 The nucleic acid fragments of the instant invention may be used to create transgenic plants in which the *Vernonia galamenensis* fatty acid modifying enzymes are present at higher levels than normal or in cell types or developmental stages in which it is not normally found. For example, when over-expressed in plant cells, the *Vernonia galamenensis* fatty acid epoxidizing enzyme may be useful for causing the biosynthesis and accumulation of epoxidized fatty acids in those cells. It is particularly useful to use the *Vernonia galamenensis* fatty acid epoxidizing enzyme gene to produce epoxidized fatty acids in the cells of the seeds of oilseed crop plants.

35 Overexpression of the *Vernonia galamenensis* fatty acid epoxidizing or desaturase enzymes may be accomplished by first constructing a chimeric gene in which the *Vernonia*

galamenensis fatty acid epoxidizing or desaturase enzyme coding region is operably linked to a promoter capable of directing expression of a gene in the desired tissues at the desired stage of development. For reasons of convenience, the chimeric gene may comprise a promoter sequence and translation leader sequence derived from the same gene. 3' non-coding sequences encoding transcription termination signals may also be provided. The instant chimeric genes may also comprise one or more introns in order to facilitate gene expression.

Plasmid vectors comprising the instant chimeric genes can then be constructed. The choice of plasmid vector is dependent upon the method that will be used to transform host plants. The skilled artisan is well aware of the genetic elements that must be present on the plasmid vector in order to successfully transform, select and propagate host cells containing the chimeric gene. The skilled artisan will also recognize that different independent transformation events will result in different levels and patterns of expression (Jones et al., (1985) *EMBO J.* 4:2411-2418; De Almeida et al., (1989) *Mol. Gen. Genetics* 218:78-86), and thus that multiple events must be screened in order to obtain lines displaying the desired expression level and pattern. Such screening may be accomplished by Southern analysis of DNA, Northern analysis of mRNA expression, Western analysis of protein expression, or phenotypic analysis.

For some applications it may be useful to direct the instant *Vernonia galamenensis* fatty acid modifying enzymes to different cellular compartments, or to facilitate its secretion from the cell. It is thus envisioned that the chimeric genes described above may be further supplemented by altering the coding sequences to encode *Vernonia galamenensis* fatty acid modifying enzymes disclosed herein with appropriate intracellular targeting sequences such as transit sequences (Keegstra, K. (1989) *Cell* 56:247-253), signal sequences or sequences encoding endoplasmic reticulum localization (Chrispeels, J.J., (1991) *Ann. Rev. Plant Phys. Plant Mol. Biol.* 42:21-53), or nuclear localization signals (Raikhel, N. (1992) *Plant Phys.* 100:1627-1632) added and/or with targeting sequences that are already present removed. While the references cited give examples of each of these, the list is not exhaustive and more targeting signals of utility may be discovered in the future.

The instant *Vernonia galamenensis* fatty acid modifying enzymes (or portions of the enzymes) may be produced in heterologous host cells, particularly in the cells of microbial hosts, and can be used to prepare antibodies to the *Vernonia galamenensis* fatty acid epoxidizing and desaturase enzymes by methods well known to those skilled in the art. The antibodies are useful for detecting the instant *Vernonia galamenensis* fatty acid modifying enzymes *in situ* in cells or *in vitro* in cell extracts. Preferred heterologous host cells for production of the instant *Vernonia galamenensis* fatty acid modifying enzymes are microbial hosts. Microbial expression systems and expression vectors containing regulatory sequences that direct high level expression of foreign proteins are well known to those skilled in the art.

Any of these could be used to construct chimeric genes for production of the instant *Vernonia galamenensis* fatty acid epoxidizing enzyme or the instant *Vernonia galamenensis* fatty acid desaturase enzyme. These chimeric genes could then be introduced into appropriate microorganisms via transformation to provide high level expression of the 5 encoded *Vernonia galamenensis* fatty acid modifying enzyme. An example of a vector for high level expression of the instant *Vernonia galamenensis* fatty acid modifying enzymes in a bacterial host is provided (Example 8).

All or a portion of the nucleic acid fragments of the instant invention may also be used as probes for genetically and physically mapping the genes that they are a part of, and as 10 markers for traits linked to expression of the instant fatty acid epoxidizing and desaturase enzymes. Such information may be useful in plant breeding in order to develop lines with desired phenotypes.

For example, the instant nucleic acid fragments may be used as restriction fragment length polymorphism (RFLP) markers. Southern blots (Maniatis) of restriction-digested 15 plant genomic DNA may be probed with the nucleic acid fragments of the instant invention. The resulting banding patterns may then be subjected to genetic analyses using computer programs such as MapMaker (Lander et al., (1987) *Genomics* 1:174-181) in order to construct a genetic map. In addition, the nucleic acid fragments of the instant invention may be used to probe Southern blots containing restriction endonuclease-treated genomic DNAs 20 of a set of individuals representing parent and progeny of a defined genetic cross. Segregation of the DNA polymorphisms is noted and used to calculate the position of the instant nucleic acid sequence in the genetic map previously obtained using this population (Botstein, D. et al., (1980) *Am.J.Hum.Genet.* 32:314-331).

The production and use of plant gene-derived probes for use in genetic mapping is 25 described in R.Bernatzky, R. and Tanksley, S.D. (1986) *Plant Mol.Biol.Reporter* 4(1):37-41. Numerous publications describe genetic mapping of specific cDNA clones using the methodology outlined above or variations thereof. For example, F2 intercross populations, backcross populations, randomly mated populations, near isogenic lines, and other sets of individuals may be used for mapping. Such methodologies are well known to those skilled 30 in the art.

Nucleic acid probes derived from the instant nucleic acid sequences may also be used for physical mapping (i.e., placement of sequences on physical maps; see Hoheisel, J. D., et al., In: *Nonmammalian Genomic Analysis: A Practical Guide*, Academic press 1996, pp. 319-346, and references cited therein).

In another embodiment, nucleic acid probes derived from the instant nucleic acid 35 sequences may be used in direct fluorescence *in situ* hybridization (FISH) mapping. Although current methods of FISH mapping favor use of large clones (several to several

hundred KB), improvements in sensitivity may allow performance of FISH mapping using shorter probes.

A variety of nucleic acid amplification-based methods of genetic and physical mapping may be carried out using the instant nucleic acid sequences. Examples include 5 allele-specific amplification, polymorphism of PCR-amplified fragments (CAPS), allele-specific ligation, nucleotide extension reactions, Radiation Hybrid Mapping and Happy Mapping. For these methods, the sequence of a nucleic acid fragment is used to design and produce primer pairs for use in the amplification reaction or in primer extension reactions. The design of such primers is well known to those skilled in the art. In methods employing 10 PCR-based genetic mapping, it may be necessary to identify DNA sequence differences between the parents of the mapping cross in the region corresponding to the instant nucleic acid sequence. This, however, is generally not necessary for mapping methods. Such information may be useful in plant breeding in order to develop lines with desired starch phenotypes.

15

EXAMPLES

The present invention is further defined in the following Examples, in which all parts and percentages are by weight and degrees are Celsius, unless otherwise stated. It should be understood that these Examples, while indicating preferred embodiments of the invention, are given by way of illustration only. From the above discussion and these Examples, one 20 skilled in the art can ascertain the essential characteristics of this invention, and without departing from the spirit and scope thereof, can make various changes and modifications of the invention to adapt it to various usages and conditions.

EXAMPLE 1

Composition of a cDNA Library; Isolation and Sequencing of cDNA Clones

25

A cDNA library representing mRNAs from developing seeds of *Vernonia galamensis* that had just begun production of vernolic acid was prepared. The library was prepared in a Uni-ZAP™ XR vector according to the manufacturer's protocol (Stratagene Cloning Systems, La Jolla, CA). Conversion of the Uni-ZAP™ XR library into a plasmid library was accomplished according to the protocol provided by Stratagene. Upon 30 conversion, cDNA inserts were contained in the plasmid vector pBluescript. cDNA inserts from randomly picked bacterial colonies containing recombinant pBluescript plasmids were amplified via polymerase chain reaction using primers specific for vector sequences flanking the inserted cDNA sequences. Amplified insert DNAs were sequenced in dye-primer sequencing reactions to generate partial cDNA sequences (expressed sequence tags or 35 "ESTs"; see Adams, M. D. et al., (1991) *Science* 252:1651). The resulting ESTs were analyzed using a Perkin Elmer Model 377 fluorescent sequencer.

EXAMPLE 2Identification and Characterization of cDNA Clones

ESTs encoding *Vernonia galamenensis* fatty acid modifying enzymes were identified by conducting BLAST (Basic Local Alignment Search Tool; Altschul, S. F., et al., (1993) 5 *J. Mol. Biol.* 215:403-410; see also www.ncbi.nlm.nih.gov/BLAST/) searches for similarity to sequences contained in the BLAST “nr” database (comprising all non-redundant GenBank CDS translations, sequences derived from the 3-dimensional structure Brookhaven Protein Data Bank, the last major release of the SWISS-PROT protein sequence database, EMBL, and DDBJ databases). The cDNA sequences obtained in Example 1 were analyzed 10 for similarity to all publicly available DNA sequences contained in the “nr” database using the BLASTN algorithm provided by the National Center for Biotechnology Information (NCBI). The DNA sequences were translated in all reading frames and compared for similarity to all publicly available protein sequences contained in the “nr” database using the BLASTX algorithm (Gish, W. and States, D. J. (1993) *Nature Genetics* 3:266-272) 15 provided by the NCBI. For convenience, the P-value (probability) of observing a match of a cDNA sequence to a sequence contained in the searched databases merely by chance as calculated by BLAST are reported herein as “pLog” values, which represent the negative of the logarithm of the reported P-value. Accordingly, the greater the pLog value, the greater the likelihood that the cDNA sequence and the BLAST “hit” represent homologous 20 proteins.

The BLASTX search using the nucleotide sequence from clone vs1.05h08 revealed similarity of the protein encoded by the cDNA to, *inter alia*, a tomato fatty acid desaturase enzyme (EMBL Accession No. X94944; pLog = 1.72) and a potato fatty acid desaturase enzyme (EMBL Accession No. X92847; pLog = 0.52). The sequence of the entire cDNA 25 insert in clone vs1.05h08 was determined and reevaluated by BLAST, yielding even higher pLog values vs. the potato fatty acid desaturase enzyme (X92847; pLog = 228.04). SEQ ID NO:1 shows the nucleotide sequence of the entire *Vernonia galamenensis* cDNA in clone vs1.05h08; the deduced amino acid sequence is shown in SEQ ID NO:2. Sequence alignments and BLAST scores and probabilities indicate that the instant nucleic acid 30 fragment encodes a *Vernonia galamenensis* fatty acid desaturase.

The BLASTX search using the nucleotide sequence of clone vs1.02c07 revealed similarity of the protein encoded by the cDNA to, *inter alia*, a potato fatty acid desaturase enzyme (EMBL Accession No. X92847; pLog = 2.08) and a soybean fatty acid desaturase enzyme (GenBank Accession No. L43921; pLog = 1.49). The sequence of the entire cDNA 35 insert in clone vs1.02c07 was determined and reevaluated by BLAST, yielding even higher pLog values vs. the potato (X92847; pLog = 156.18) and soybean fatty acid desaturase enzymes (L43921; pLog = 152.18). SEQ ID NO:3 shows the nucleotide sequence of the

entire *Vernonia galamenensis* cDNA; the deduced amino acid sequence is shown in SEQ ID NO:4.

The deduced amino acid sequences from cDNA clones vs1.05h08 and vs1.02c07 were compared to the deduced amino acid sequences encoding (i) a known fatty acid desaturase from soybean (World Patent Publication No. WO94/11516) and (ii) a fatty acid hydroxylase from castor bean (van de Loo, F. J. et al. (1995) *Proc. Natl. Acad. Sci. U.S.A.* 92 (15):6743-6747) using the multiple sequence comparison program "Megalign" from the Lasargene™ software package (DNASTAR Inc., Madison, WI). The aligned sequences are shown in Figure 1. All four sequences are related by eight very highly conserved residues that are apparently part of the binding site for the two iron cluster that is required in the active site of enzymes in this class (Shanklin, J. et al. (1994) *Biochemistry* 33:12787-12793). The cDNA insert in clone vs1.05h08 is about 70% similar to the known fatty acid desaturase from soybean, while the cDNA insert in vs1.02c07 is only 53.8% similar to this soybean fatty acid desaturase. This degree of divergence is similar to that observed between the fatty acid hydroxylase from castor bean and the fatty acid desaturases. Thus, changes in a comparatively small number of amino acid residues in conserved regions of the protein are sufficient to alter the activity in this class of enzymes from one of introducing a double bond (i.e., a desaturase) to one of introducing an hydroxyl group (i.e., a hydroxylase).

The sequence of clone vs1.02c07 is also quite divergent from the castor bean fatty acid hydroxylase, sharing only 52.5% similar residues. The sequence of vs1.02c07 is therefore unique, but nonetheless related, to the class of enzymes which is known to contain enzymes capable of producing different modifications in the acyl chains of fatty acids. Sequence alignments, BLAST scores and probabilities and experimental data demonstrating a tissue-specific expression pattern (see Example 3) for the gene encoded by the cDNA clone vs1.02c07 indicate that the instant nucleic acid fragment comprising this clone encodes a *Vernonia galamenensis* fatty acid epoxidizing enzyme.

EXAMPLE 3

Tissue Specific Expression of the Fatty Acid Epoxidase

Expression of enzymes which produce unusual fatty acids has been shown to be specific for the storage organs in which these fatty acids are found (see World Patent Publication No. WO94/11516). Northern analysis of mRNA from developing leaves of *Vernonia galamenensis* was performed using the cDNA insert in vs1.02c07 as a probe. Messenger RNA from leaves that were near full expansion (by comparison to other leaves just lower on the main stem) were removed and mRNA was prepared by standard methods well known in the art. The leaf mRNA, and a remaining sample of the developing seed mRNA that was used as template for the cDNA library from which the instant cDNA clones were obtained, was separated by denaturing agarose gel electrophoresis and blotted to a nylon membrane for hybridization to the probe.

The ^{32}P -labeled probe based on the cDNA insert in vs1.02c07 was prepared by PCR amplification of the coding region of the cDNA using the nucleotide described in SEQ ID NO:7 as the 5' end primer and the nucleotide described in SEQ ID NO:8 as the 3' end primer. The product from PCR amplification was purified by isolation from an agarose gel and used 5 as template for random primed labeling. The portion of cDNA clone vs1.05h08 encoding the peptide was similarly amplified using the nucleotides described in SEQ ID NOs:9 and 10 as the 5' and 3' primers, respectively, and the purified product used as probe template.

Both probes were hybridized to the Northern blots at 62°C in 0.2X SSC overnight. Excess probe was removed by washing under the same stringency conditions and the blot 10 was placed on photographic film for development.

The lane containing the seed mRNA produced an intense hybridization signal at about 1.9kD when the coding region of vs1.02c07 was used as the probe, while no signal was visible in the lane containing the leaf-derived mRNA. Further exposure of the blot, such that the seed-derived signal was highly over exposed, still did not result in a visible signal in the 15 lane containing the leaf-derived message. The coding region of vs1.05h08 hybridized to message from both developing seeds and from leaves.

The cDNA insert in vs1.02c07 is therefore not expressed in a *Vernonia galamenensis* tissue that does not produce epoxidized fatty acids, but is highly expressed in a tissue that does produce the modified fatty acid. The tissue-specific nature of its expression, its 20 relationship to a known class of fatty acid modifying enzymes, and its divergence from enzymes in that class whose catalytic function has been demonstrated, all indicate that the cDNA insert in vs1.02c07 encodes the fatty acid epoxidizing enzyme from *Vernonia galamenensis*.

EXAMPLE 4

Expression of Chimeric Genes in Monocot Cells

The oil storing tissues of most grass seeds are the embryo and its attending tissues the scutellum and to some extent the aleurone. Promoter sequences such as those controlling expression of the storage proteins Globulin 1 (Belanger, S.C. and Kriz, A. L. (1989) *Plant Physiol.* 91:636-643) and Globulin 2 (Wallace, N.H. and Kriz, A.L. (1991) *Plant Physiol.* 95:973-975) are appropriate for the expression of chimeric genes in these tissues.

A chimeric gene comprising a cDNA encoding a *Vernonia galamenensis* fatty acid epoxidizing enzyme in sense orientation with respect to the maize Globulin 2 promoter that is located 5' to the cDNA fragment, and the Globulin 2 3' end that is located 3' to the cDNA fragment, can be constructed. The cDNA fragment of this gene may be generated by 35 polymerase chain reaction (PCR) of the cDNA clone using appropriate oligonucleotide primers. Cloning sites can be incorporated into the oligonucleotides to provide proper orientation of the DNA fragment when inserted into the correctly designed expression vector.

Such expression vectors should include genetic sequence elements conferring an origin of replication for the plasmid in its host, a gene capable of conferring a selectable trait such as autotrophy or antibiotic tolerance to the host cell carrying the plasmid, and the promoter sequences for expression of desired genes in host plant cells. Further design features may 5 include unique restriction endonuclease recognition sites between the elements of the plant gene promoter elements to allow convenient introduction genes to be controlled by those elements.

The chimeric genes constructed as above can then be introduced into corn cells by the following procedure. Immature corn embryos can be dissected from developing caryopses 10 derived from crosses of the inbred corn lines H99 and LH132. The embryos are isolated 10 to 11 days after pollination when they are 1.0 to 1.5 mm long. The embryos are then placed with the axis-side facing down and in contact with agarose-solidified N6 medium (Chu et al., (1975) *Sci. Sin. Peking* 18:659-668). The embryos are kept in the dark at 27°C. Friable embryogenic callus consisting of undifferentiated masses of cells with somatic 15 proembryoids and embryoids borne on suspensor structures proliferates from the scutellum of these immature embryos. The embryogenic callus isolated from the primary explant can be cultured on N6 medium and sub-cultured on this medium every 2 to 3 weeks.

The plasmid, p35S/Ac (obtained from Dr. Peter Eckes, Hoechst Ag, Frankfurt, 20 Germany) may be used in transformation experiments in order to provide for a selectable marker. This plasmid contains the *Pat* gene (see European Patent Publication 0 242 236) which encodes phosphinothricin acetyl transferase (PAT). The enzyme PAT confers 25 resistance to herbicidal glutamine synthetase inhibitors such as phosphinothricin. The *pat* gene in p35S/Ac is under the control of the 35S promoter from Cauliflower Mosaic Virus (Odell et al. (1985) *Nature* 313:810-812) and the 3' region of the nopaline synthase gene from the T-DNA of the Ti plasmid of *Agrobacterium tumefaciens*.

The particle bombardment method (Klein et al., (1987) *Nature* 327:70-73) may be used to transfer genes to the callus culture cells. According to this method, gold particles (1 µm in diameter) are coated with DNA using the following technique. Ten µg of plasmid 30 DNAs are added to 50 µL of a suspension of gold particles (60 mg per mL). Calcium chloride (50 µL of a 2.5 M solution) and spermidine free base (20 µL of a 1.0 M solution) are added to the particles. The suspension is vortexed during the addition of these solutions. After 10 minutes, the tubes are briefly centrifuged (5 sec at 15,000 rpm) and the supernatant removed. The particles are resuspended in 200 µL of absolute ethanol, centrifuged again and the supernatant removed. The ethanol rinse is performed again and the particles 35 resuspended in a final volume of 30 µL of ethanol. An aliquot (5 µL) of the DNA-coated gold particles can be placed in the center of a Kapton™ flying disc (Bio-Rad Labs). The particles are then accelerated into the corn tissue with a Biolistic™ PDS-1000/He (Bio-Rad

Instruments, Hercules CA), using a helium pressure of 1000 psi, a gap distance of 0.5 cm and a flying distance of 1.0 cm.

For bombardment, the embryogenic tissue is placed on filter paper over agarose-solidified N6 medium. The tissue is arranged as a thin lawn and covered a circular area of about 5 cm in diameter. The petri dish containing the tissue can be placed in the chamber of the PDS-1000/He approximately 8 cm from the stopping screen. The air in the chamber is then evacuated to a vacuum of 28 inches of Hg. The macrocarrier is accelerated with a helium shock wave using a rupture membrane that bursts when the He pressure in the shock tube reaches 1000 psi.

5 Seven days after bombardment the tissue can be transferred to N6 medium that contains glufosinate (2 mg per liter) and lacks casein or proline. The tissue continues to grow slowly on this medium. After an additional 2 weeks the tissue can be transferred to fresh N6 medium containing glufosinate. After 6 weeks, areas of about 1 cm in diameter of actively growing callus can be identified on some of the plates containing the glufosinate-supplemented medium. These calli may continue to grow when sub-cultured on the 10 selective medium.

15 Plants can be regenerated from the transgenic callus by first transferring clusters of tissue to N6 medium supplemented with 0.2 mg per liter of 2,4-D. After two weeks the tissue can be transferred to regeneration medium (Fromm et al., (1990) *Bio/Technology* 8:833-839).

20 EXAMPLE 5
Expression of Chimeric Genes in Dicot Cells

The *Vernonia galamenensis* fatty acid epoxidizing enzyme was expressed in cells of two dicots that normally produce storage lipid by the construction of appropriate chimeric genes followed by stable introduction of those genes into the host plants.

25 Soybean embryo expression

A plasmid designated pKS18HH containing chimeric genes to afford expression of Hygromycin B Phosphotransferase in certain bacteria and in plant cells was constructed from the following genetic elements: a) T7 Promoter and Shine-Delgarno

30 sequence/Hygromycin B Phosphotransferase (HPT)/T7 Terminator Sequence, b) 35S Promoter from cauliflower mosaic virus (CaMV)/Hygromycin B Phosphotransferase (HPT)/Nopaline Synthase (NOS 3' from *Agrobacterium tumefaciens* T-DNA, and c) pSP72 plasmid vector (Promega Biotech) with beta-lactamase coding region (ampicillin resistance gene) removed. The Hygromycin B Phosphotransferase gene was obtained by PCR 35 amplification of the *Klebsiella* derived plasmid pJR225 (Gritz, L. and Davies, J. (1983) *Gene* 25(2-3):179-88) in *E. coli* strain W677. Starting with the pSP72 vector, the elements were assembled into a single plasmid using standard cloning methods (Maniatis).

Plasmid pKS18HH thus contains the T7 promoter/HPT/T7 terminator cassette for expression of the HPT enzyme in certain strains of *E. coli*, such as NovaBlue(DE3) (Novagen), that are lysogenic for lambda DE3 (which carries the T7 RNA Polymerase gene under lacUV5 control). Plasmid pKS18HH also contains the 35S/HPT/NOS cassette for 5 constitutive expression of the HPT enzyme in plants, such as soybean. These two expression systems allow selection for growth in the presence of hygromycin to be used as a means of identifying cells that contain the plasmid in both bacterial and plant systems. Plasmid pKS18HH also contains three unique restriction endonuclease sites suitable for the cloning other chimeric genes into this vector.

10 A plasmid for expression of the cDNA encoding the *Vernonia galamenensis* fatty acid epoxidizing enzyme under control of the soybean beta-conglycinin promoter (Beachy et al. (1985) *EMBO J.* 4:3047-3053) was constructed. The construction of this vector was facilitated by the use of plasmids pCW109 and pML18, both of which have been described (see World Patent Publication No. WO94/11516).

15 A unique Not I site was introduced into the cloning region between the beta-conglycinin promoter and the phaseolin 3' end in pCW109 by digestion with Nco I and Xba I followed by removal of the single stranded DNA ends with mung bean exonuclease. Not I linkers (New England Biolabs Inc., cat. No. NEB 1125) were ligated into the linearized plasmid to produce plasmid pAW35. The single Not I site in pML18 was destroyed by 20 digestion with Not I, filling in the single stranded ends with dNTP's and Klenow fragment followed by re-ligation of the linearized plasmid. The modified pML18 was then digested with Hind III and treated with calf intestinal phosphatase.

25 The beta-conglycinin/Not I/phaseolin expression cassette in pAW35 was removed by digestion with Hind III and the 1.79 kB fragment was isolated by agarose gel electrophoresis. The isolated fragment was ligated into the modified and linearized pML18 construction described above. A clone with the desired orientation was identified by digestion with Not I and Xba I to release a 1.08 kB fragment indicating that the orientation of the beta-conglycinin transcription unit was the same as the selectable marker transcription unit. The resulting plasmid was given the name pBS19.

30 Hind III is one of the unique cloning sites available in pKS18HH. To assemble the final expression cassette, pBS19 and pKS18HH were both digested with Hind III. The beta-conglycinin containing fragment from pBS19 was isolated by gel electrophoresis and ligated into the digested pKS18HH which had been treated with calf alkaline phosphatase. The resulting plasmid was named pRB20.

35 The PCR product amplified from clone vs1.02c07 (described in Example 3 above) was digested with Not I to cleave the Not I sites designed into the PCR primers. Plasmid pRB20 was also Not I digested. After phosphatase treatment of the linearized pRB20, the Not I digested vs1.02c07 product was ligated into pRB20 and the ligation mixture used to

transform *E. coli* strain DE3. Colonies were selected and grown in liquid media for preparation of plasmid DNA. Digestion of the plasmid DNA with Xmn I released a fragment of 0.4 kB when the coding sequence of vs1.02c07 was oriented in the sense direction relative to the beta-conglycinin promoter. The selected clone was designated 5 pRVF7 and the restriction endonuclease map of the plasmid is shown in Figure 2. The *E. coli* cell line containing pRVF7 was grown and plasmid DNA isolated to produce DNA for stable transformation of soybean.

Soybean embryos were transformed with the expression vector pRVF7. Media designations and recipes are given in Table 1 below. To induce somatic embryos, 10 cotyledons 3-5 mm in length dissected from surface sterilized, immature seeds of the soybean cultivar A2872, were cultured in the light or dark at 26°C on agar medium SB1 for 6-10 weeks. Somatic embryos which produced secondary embryos were excised and placed into liquid medium SB 172. After repeated selection for clusters of somatic embryos which multiplied as early, globular staged embryos, the suspensions were maintained as described 15 below.

Soybean embryogenic suspension cultures were maintained in 35 mL of SB 172 media on a rotary shaker, 150 rpm, at 26°C with fluorescent lights on a 16:8 hour day/night schedule. Cultures were subcultured every two weeks by inoculating approximately 35 mg of tissue into 35 mL of SB 172 medium.

20 Soybean embryogenic suspension cultures were transformed by the method of particle gun bombardment (Kline et al. (1987) *Nature (London)* 327:70; U.S. Patent No. 4,945,050) using a Du Pont Biolistic™ PDS1000/HE instrument (helium retrofit) can.

To 50 mL of a 60 mg/mL 1 mm gold particle suspension was added (in order): 5 mL of plasmid pRVF7 DNA (1 mg/mL), 20 ml spermidine (0.1 M), and 50 mL CaCl2 (2.5 M). 25 The particle preparation was agitated for three minutes, spun in a microfuge for 10 seconds and the supernatant removed. The DNA-coated particles were washed once in 400 mL 70% ethanol and resuspended in 40 mL of anhydrous ethanol. The DNA/particle suspension was sonicated three times for one second each. Five mL of the DNA-coated gold particles were then loaded on each macro carrier disk.

30 Approximately 300-400 mg of a two-week-old suspension culture was placed in an empty 60x15 mm petri dish and the residual liquid removed from the tissue with a pipette. For each transformation experiment, approximately 5-10 plates of tissue were bombarded. Membrane rupture pressure was set at 1100 psi and the chamber was evacuated to a vacuum of 28 inches mercury. The tissue was placed approximately 3.5 inches away from the 35 retaining screen and bombarded three times. Following bombardment, the tissue was divided in half and placed back into liquid and cultured as described above.

Three to four days post bombardment, the liquid media was exchanged with fresh media, and six to eleven days post bombardment with fresh SB 172 media containing

50 mg/L hygromycin. This selective media was refreshed weekly. Four to six weeks post bombardment, green, transformed tissue was observed growing from untransformed, necrotic embryogenic clusters. Isolated green tissue was removed and inoculated into individual cells of six cell well plates. Each well contained six ml of SB 172 media with no 5 hygromycin. Cultures were maintained for two to six weeks with media change every two weeks. Surviving cultures are treated as independent transformation events and were returned to 35 mL SB 172 with hygromycin in culture flasks for tissue increase for four to six weeks with media changes every two weeks.

10 Embryonic clusters were removed from liquid culture and placed on solid agar SB166 media for one week then subcultured to SB103 for 3 weeks. Individual, maturing embryos were removed for fatty acid analysis as described in Example 7 at this point.

15 To regenerate fertile soybean plants, embryos from selected lines were desiccated by placing embryos into a small, empty petri plate that was placed inside a sealed, larger petri plate containing SB103 media. Partially desiccated embryos that were yellow in color were returned to petri plates containing either SB71-3 or SB71-4 media and allowed to germinate. Embryos that germinated and produced roots and leaves were transferred to sterile soil and grown to maturity for seed collection.

Table 1. Transformation Media

SB172 (per liter)	1 bottle Duchefa salts (MS medium-Finer modification)* 1 ml B5 vitamins 1 ml 2,4-D stock 60 g sucrose 0.667 g asparagine pH 5.7
SB1 (per liter)	1 pkg MS salts (Gibco/BRL - premade formulation) 1 ml B5 vitamins 60 g sucrose 4 ml 2,4-D stock pH 5.8 0.8% agar
SB166 (per liter)	1 pkg MS salts (Gibco/BRL - premade formulation) 1 ml B5 vitamins 60 g maltose 750 mg MgCl ₂ hexahydrate 5 g activated charcoal pH 5.7 2 g gelrite

SB103 (per liter)	1 pkg MS salts (Gibco/BRL - premade formulation) 1 ml B5 vitamins 60 g maltose 750 mg MgCl ₂ hexahydrate pH 5.7 2 g gelrite*
SB 148 (per liter)	1 pkg MS salts (Gibco/BRL - premade formulation) 1 ml B5 vitamins 60 g maltose pH 5.7 7 g agarose
SB 71-3 (per liter)	1 bottle B5 salts w/ sucrose (Gibco/BRL - premade formulation) 750 mg MgCl ₂ hexahydrate pH 5.7 5 g TC agar
SB 71-4 (per liter)	1 bottle B5 salts w/ sucrose (Gibco/BRL - premade formulation) pH 5.7 5 g TC agar
2,4-D stock	10 mg/ml
B5 vitamins	1000X stock - (per 100 ml) 10 g myo-inositol 100 mg nicotinic acid 100 mg pyridoxine HCl 1 g thiamine

*Duchefa Biochemie B.V., Izaak Enschedeweg 40, NL-2031 CS Haarlem, Netherland

EXAMPLE 6

5

Seed Specific Expression of the Vernonia Fatty Acid Epoxidase in *Arabidopsis thaliana*

Arabidopsis thaliana was transformed with a vector designed to allow seed specific expression of the vernonia sequence vs1.02c07 using the *in planta* transformation procedure described by Bechtold et al. ((1993) *CR Acad Sci Paris* 316:1194-1199). The expression vector was constructed using two existing plasmids. Plasmid pML2 contained the cDNA sequence for a mutant form of the enzyme acetolactate synthase from *Arabidopsis* under the control of the 35S promoter. Transfer to plants confers resistance to the sulfonyl urea herbicide chlorsufuron. Plasmid pIMC01 contains the 5' and 3' regulatory regions surrounding the *Brassica* storage protein Napin with added restriction endonuclease sites between the 5' and 3' regulatory regions. Its construction is described in WO94/11516 as is

the Agrobacterium binary vector pZS94K. NcoI and SalI sites were introduced on the 5' and 3' ends of vs1.02c07 by PCR using primers that contain the recognition sequences for those enzymes 5' and 3' respectively to the target sequences at the ends of vs1.02c07. The PCR product was digested with NcoI and SalI and ligated into pIMC01 that had also been 5 digested with NcoI and SalI.

Seed from the primary transformants was planted in flats, cold treated before germination and the seedlings were sprayed with 50 mg/L chlorsulfuron. Surviving seedlings were transplanted, taken to maturity and seeds were analyzed for the presence of the 12, 13 epoxide containing vernolic acid as described below. Seeds from two lines that produced 10 seeds segregating for production of vernolic acid were planted and the seedlings were sprayed with 200 mg/L chlorsulfuron. The surviving seedlings were again grown to maturity and the resulting seeds analyzed for the presence of vernolic acid.

EXAMPLE 7

Analysis of Vernolic Acid in Transgenic Plant Tissues

15 The *Arabidopsis* transformation procedure resulted in many apparent escape events as only one in twenty plants chosen as possible transformants had a seed fatty acid profile that included the putative vernolic acid. That one event accumulated the new fatty acid to about 1% of the total fatty acids in individual seeds from the transformant. This accumulation level was sufficient to allow positive identification of the putative vernolic acid derivative by 20 GC-mass spectrometry. GC-MS was done first on the total fatty acids from a single seed which was shown by GC analysis to contain the putative vernolic methyl ester by the method described in Hitz et al. ((1994) *Plant Physiol.* 105:635-641). The mass spectra of the major peak in the methyl esters obtained from mature *Vernonia galemenensis* had a weak molecular ion at M/e+ 310 and a slightly more prominent signal at (M-31) of 279. This is a 25 common initial loss of the OCH₃ from fatty acid methyl esters. While the transgenic *arabidopsis* seed did not provide sufficient mass spectral signal to identify the molecular ion in the putative vernolic acid methyl ester peak the signal at M/e+ 279 was present and the remainder of the spectra was identical to the main peak from *vernonia*.

30 Single seed analysis of twelve seeds from one of the *Arabidopsis* plants selected by spraying with chlorsulfuron in the second generation after transformation all contained the putative vernolic acid at about 1.5% of the total fatty acids. The plant is apparently homozygous for the transgene.

35 Methyl esterification of the fatty acids from *Vernonia* under acid conditions results in a slow methanolysis of the 12, 13 epoxide to a mixture of the 12-OH, 13- OCH₃ and the 12- OCH₃, 13-OH methyl esters. Bulk seed from the selected, homozygous transformant was treated with 1% H₂SO₄ in methanol at reflux for 4 h and the resulting fatty acid methyl esters run on GC-MS as above. The larger of the two peaks that were unique to the transformed *arabidopsis* fatty acids had a spectra characteristic of the 12-OH, 13-OCH₃

methyl ester derivative of the methanolysis product of vernolic acid: a prominent fragment at M+/e of 227 corresponding to the portion of the molecule from the carboxyl end to the 12 carbon and the base ion at M+/e 115 corresponding to the fragment from carbons 13 through 18. The other peak produced a spectra characteristic of the 13- OCH₃, 12-OH methyl ester: 5 the M+/e fragment at 241 coresponding to the split between carbons 12 and 13 with the 12 carbon carrying the methoxy group and the base ion at M+/e 145 from the split between carbons 11 and 12.

10 The results from the *Arabadopsis* transformations prove that the enzyme product encoded by SEQ ID NO:3 is capable of catalyzing the production of vernolic acid when transgenically introduced into a plant that does not normally produce or accumulate vernolic acid.

15 Fatty acids in somatic soybean embryos were directly transesterified in methanol containing 1% sodium methoxide. The fatty acid methyl esters were extracted into heptane and a portion of the heptane extract was seperated by gas liquid chromatography as described in Hitz et al. ((1994) *Plant Physiol* 105:635-641). Ten soybean embryos from each of 8 transgenic lines were assayed individually and the results are given in Table 2.

Table 2. Individual fatty acids as a per cent of total fatty acid in single somatic embryos obtained after transformation with plasmid pRFV7.

Line#	Sample	16:0	18:0	18:1	18:2	18:3	Vern
2065/3/6/1	2065361.0	14.5	2.5	8.8	58.7	15.1	0.4
	2065361.1	12.9	3.1	13.9	56.3	12.3	1.5
	2065361.2	15.2	3.7	12.2	51.7	16.2	1.1
	2065361.3	0.0	4.5	18.2	55.3	19.0	2.9
	2065361.4	17.0	4.3	9.4	49.3	18.2	1.8
	2065361.5	0.0	6.1	14.5	59.0	19.5	0.9
	2065361.6	16.2	5.3	12.9	48.4	15.3	1.9
	2065361.7	14.9	4.9	10.0	51.3	15.5	3.5
	2065361.8	14.0	3.6	12.8	55.1	13.3	1.3
	2065361.9	16.6	4.5	11.5	50.0	15.5	1.8
2065/3/6/3	2065363.0	13.8	4.1	13.8	34.1	28.5	5.8
	2065363.1	13.7	3.3	11.7	50.1	20.1	1.1
	2065363.2	16.1	4.3	10.1	39.2	25.9	4.4
	2065363.3	13.7	4.5	11.0	50.3	18.5	2.0
	2065363.4	11.8	3.7	14.4	54.0	14.4	1.7
	2065363.5	18.7	4.9	8.0	40.6	27.8	0.0
	2065363.6	13.0	2.6	10.1	54.2	17.9	2.3
	2065363.7	13.0	3.1	11.2	55.7	16.1	0.9
	2065363.8	19.5	4.3	7.9	43.9	24.4	0.0
	2065363.9	16.0	3.5	7.5	49.1	23.3	0.7

Line#	Sample	16:0	18:0	18:1	18:2	18:3	Vern
2065/3/11/1	20653111.0	15.3	2.5	8.3	53.6	20.2	0.0
	20653111.1	15.6	5.4	10.3	49.3	19.1	0.3
	20653111.2	15.9	3.5	10.0	46.7	21.4	2.5
	20653111.3	12.3	2.7	14.7	57.8	12.5	0.0
	20653111.4	16.6	4.4	8.2	51.3	18.0	1.5
	20653111.5	14.8	3.2	8.5	51.2	20.3	2.0
	20653111.6	13.9	2.9	11.0	58.4	13.9	0.0
	20653111.7	12.9	3.4	9.6	58.5	15.6	0.0
	20653111.8	14.4	3.9	6.5	54.0	19.2	2.0
	20653111.9	13.6	3.0	11.6	58.3	13.5	0.0
2065/3/11/2	20653112.0	12.8	3.6	11.4	53.8	18.2	0.0
	20653112.1	11.0	2.8	15.4	55.5	15.3	0.0
	20653112.2	16.3	4.6	10.8	45.8	20.0	2.6
	20653112.3	12.3	3.3	11.3	52.5	19.6	1.1
	20653112.4	15.8	4.0	6.6	34.1	30.5	8.9
	20653112.5	17.3	4.5	9.2	35.9	28.4	4.8
	20653112.6	15.5	3.4	9.8	40.5	27.4	3.4
	20653112.7	16.3	4.1	8.2	37.4	28.1	6.0
	20653112.8	17.7	4.8	8.4	34.7	27.9	6.4
	20653112.9	17.6	3.7	6.2	36.6	27.7	8.3
2092/1/1 22-Oct	2092-1-1-1	12.8	4.6	14.2	54.7	13.1	0.0
	-2.0	11.5	3.7	13.0	55.9	15.4	0.0
	-3.0	11.4	3.4	11.8	59.8	13.2	0.0
	-4.0	17.9	3.8	3.2	32.7	41.3	0.0
	-5.0	13.2	3.3	11.3	56.2	15.5	0.0
	-6.0	13.1	3.0	11.3	58.1	14.1	0.0
	-7.0	16.9	3.5	5.6	48.7	24.5	0.0
	-8.0	14.0	3.7	7.1	57.9	16.6	0.0
	-9.0	14.1	3.2	7.5	49.8	24.6	0.3
	-10.0	16.8	3.6	4.9	43.0	30.6	0.0
2092/2/1	2092-2-1-1	18.1	4.2	7.4	38.6	30.1	0.8
	-2.0	17.4	3.8	3.4	46.9	27.4	0.0
	-3.0	13.0	3.4	12.6	57.3	13.0	0.1
	-4.0	12.7	3.6	13.3	56.8	13.0	0.1
	-5.0	14.0	4.5	9.4	50.9	19.9	0.3
	-6.0	13.4	2.8	11.8	59.3	11.7	0.5
	-7.0	15.8	3.3	8.8	55.2	15.9	0.2
	-8.0	13.8	3.5	10.7	56.0	15.5	0.0
	-9.0	18.8	3.9	8.7	43.1	24.8	0.0
	-10.0	14.1	2.7	10.2	52.1	19.6	0.6

Line#	Sample	16:0	18:0	18:1	18:2	18:3	Vern
2092/2/2	2092-2-2-1	13.0	5.0	11.7	54.6	14.9	0.0
	-2.0	16.2	4.4	4.3	56.5	18.1	0.0
	-3.0	15.1	3.5	8.4	48.9	23.3	0.0
	-4.0	15.5	4.1	5.3	56.0	18.5	0.0
	-5.0	21.6	5.4	8.4	33.7	30.1	0.0
	-6.0	15.6	5.1	9.2	50.1	19.2	0.0
	-7.0	15.4	5.1	8.2	52.0	18.6	0.0
	-8.0	14.0	5.0	9.4	55.6	15.1	0.0
	-9.0	13.7	4.3	9.7	57.6	14.2	0.0
	-10.0	12.4	3.5	9.1	58.0	16.4	0.0
2092/3/1	2092-3-1-1	13.5	3.2	11.2	57.0	14.5	0.0
	-2.0	15.6	3.8	7.2	57.4	15.2	0.0
	-3.0	12.9	2.9	12.4	57.1	14.0	0.0
	-4.0	12.8	2.5	9.9	60.4	13.9	0.0
	-5.0	16.6	3.9	6.8	52.2	19.7	0.0
	-6.0	0.0	5.0	4.1	89.9	0.0	0.0
	-7.0	13.3	3.7	7.5	55.1	19.7	0.0
	-8.0	12.6	4.7	14.7	55.1	12.2	0.0
	-9.0	12.1	3.3	13.4	57.4	13.4	0.0
	-10.0	16.4	3.9	8.9	43.1	26.0	0.0

5 A fatty acid identified by retention time as vernolic acid methyl ester was found in five of the eight lines. The amount of the fatty acid varied from one transformation event to another and to a lesser extent between embryos within an event. The maximum amount of vernolic acid accumulated was about 8% of the total fatty acids.

Accumulation of Vernolic Acid in Soybean Seeds

10 A total of 20 plants were recovered from the three transformation events from bombardment number 2065 described above, 14 plants were recovered from bombardment 2092, and a further 10 plants were recovered from bombardment 798 that was not analyzed at the somatic embryo stage.

15 Since seeds from the primary transformant are genetically segregating for the transgene in the first generation after transformation, single seeds from each plant were analyzed to estimate the segregation ratio and the possible phenotype of the homozygous seed. A small piece of cotyledon tissue was cut from each of 15 single seeds from each plant. The fatty acids in the sample were derivatized and the fatty acid profile determined by GLC as described above. If care is taken to avoid the seed embryo during sampling the remaining seed can be germinated. This seed ship method accurately predicts the fatty acid profile of the whole seed and allows recovery of a plant from a seed of known phenotype.

20 Only three of the plants recovered from bombardment 2092 accumulated any vernolic acid and those accumulated less than 1% of the total seed fatty acids as vernolic. Sixteen of the 20 plants recovered from bombardment 2065 were accumulating the epoxy fatty acid in the segregating seeds. The maximum accumulation was about 9% of total fatty acids. All

ten plants from bombardment 798 accumulated vernolic acid and a total of 165 single seeds were analyzed by the seed chip method to recover putative homozygotes for advancement of the population. The results are shown as a class histogram in Figure 3. The number of seeds which were did not accumulate a significant amount of vernolic acid suggests that the 5 phenotype follows a single gene segregation pattern. If that is the case then the population of seeds comprising the upper 25% in terms of accumulation should contain the seeds homozygous for the transgene. That group is quite broad in this population but centers on about 7% vernolic acid (Figure 3).

Seeds from within the upper 25% of accumulators and from the population of non-10 accumulators were planted. Leaf tissue for DNA extraction was taken from vegetative plants from the population derived from seeds that were accumulating vernolic acid and those that were not. Southern analysis revealed a single hybridizing band of about 3 kB in size from DNA digested with Hind III in DNA from plants derived from accumulting seeds but no corresponding band from non-accumulators. The transgenic event appears to be a 15 single transgene insertion. Seeds representing this transformation event have been deposited with the American Type Culture Collection and have been assigned ATCC Accession No. _____.

Single seeds from plants that appeared to be homozygous for the transgene in pRVF7 20 were analyzed for fatty acid profile at maturity. All seeds accumulated vernolic acid with a mean content of 4.3% and a standard deviation of 1.2% between single seeds. Seeds from plants found not to contain the transgene by Southern analysis did not accumulate vernolic acid.

In soybean as in *Arabidopsis*, the gene sequence in SEQ ID NO:3 is capable of 25 conferring the ability to accumulate vernolic acid on seeds of plants when those plants are transformed with a chimeric gene containing the sequence under the control of a suitable, seed specific promoter.

EXAMPLE 8

Expression of Chimeric Genes in Microbial Cells

The cDNAs encoding the instant *Vernonia galamenensis* fatty acid epoxidizing or 30 desaturase enzymes can be inserted into the T7 *E. coli* expression vector pET24d (Novagen). For example, plasmid DNA containing a cDNA may be appropriately digested to release a nucleic acid fragment encoding the *Vernonia galamenensis* fatty acid epoxidizing enzyme. This fragment may then be purified on a 1% NuSieve GTG™ low melting agarose gel (FMC). Buffer and agarose contain 10 µg/ml ethidium bromide for visualization of the 35 DNA fragment. The fragment can then be purified from the agarose gel by digestion with GELase™ (Epicentre Technologies) according to the manufacturer's instructions, ethanol precipitated, dried and resuspended in 20 µL of water. Appropriate oligonucleotide adapters may be ligated to the fragment using T4 DNA ligase (New England Biolabs, Beverly, MA).

The fragment containing the ligated adapters can be purified from the excess adapters using low melting agarose as described above. The vector pET24d is digested, dephosphorylated with alkaline phosphatase (NEB) and deproteinized with phenol/chloroform as described above. The prepared vector pET24d and fragment can then be ligated at 16°C for 15 hours 5 followed by transformation into DH5 electrocompetent cells (GIBCO BRL). Transformants can be selected on agar plates containing 2xYT media and 50 µg/mL kanamycin. Transformants containing the gene are then screened for the correct orientation with respect to pET24d T7 promoter by restriction enzyme analysis.

Clones in the correct orientation with respect to the T7 promoter can be transformed 10 into BL21(DE3) competent cells (Novagen) and selected on 2xYT agar plates containing 50 µg/ml kanamycin. A colony arising from this transformation construct can be grown overnight at 30°C in 2xYT media with 50 µg/mL kanamycin. The culture is then diluted two fold with fresh media, allowed to re-grow for 1 h, and induced by adding isopropyl-thiogalactopyranoside to 1 mM final concentration. Cells are then harvested by 15 centrifugation after 3 h and re-suspended in 50 µL of 50 mM Tris-HCl at pH 8.0 containing 0.1 mM DTT and 0.2 mM phenyl methylsulfonyl fluoride. A small amount of 1 mm glass beads can be added and the mixture sonicated 3 times for about 5 seconds each time with a microprobe sonicator. The mixture is centrifuged and the protein concentration of the supernatant determined. One µg of protein from the soluble fraction of the culture can be 20 separated by SDS-polyacrylamide gel electrophoresis. Gels can be observed for protein bands migrating at the expected molecular weight.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:
(A) ADDRESSEE: E. I. DU PONT DE NEMOURS AND COMPANY
(B) STREET: 1007 MARKET STREET
(C) CITY: WILMINGTON
(D) STATE: DELAWARE
(E) COUNTRY: USA
(F) ZIP: 19898
(G) TELEPHONE: 302-992-4926
(H) TELEFAX: 302-773-0164
(I) TELEX: 6717325

(ii) TITLE OF INVENTION: FATTY ACID MODIFYING ENZYMES FROM
DEVELOPING SEEDS OF VERNONIA
GALAMENENSIS

(iii) NUMBER OF SEQUENCES: 10

(iv) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: DISKETTE, 3.50 INCH
(B) COMPUTER: IBM PC COMPATIBLE
(C) OPERATING SYSTEM: MICROSOFT WINDOWS 95
(D) SOFTWARE: MICROSOFT WORD VERSION 7.0A

(v) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(vi) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: 08/872,302
(B) FILING DATE: JUNE 11, 1997

(vii) ATTORNEY/AGENT INFORMATION:
(A) NAME: MAJARIAN, WILLIAM R.
(B) REGISTRATION NUMBER: 41,173
(C) REFERENCE/DOCKET NUMBER: BB-1084-A

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1476 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION: 134..1279

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

GGCACGAGCT TCGCAGGCAC AGAGAAGGAA ATTGAGCGAT TAATCGCTTC TCCGAAGTGG	60
TTGTTTCTCC AGTGCAAACC CTAGGACTCC GTATATCGAT CGAATTAGGT TGAAGTGTCT	120
CCAGAACAAAC AAA ATG GGA GCA GGA GGG TGC ATG TCT GCC TCC GAG ACA Met Gly Ala Gly Gly Cys Met Ser Ala Ser Glu Thr	169
1 5 10	
AAA ACA CAA CAA AAG AAC CCT ATC GAG CGA GTC CCT TAT GCA AAA CCT Lys Thr Gln Gln Lys Asn Pro Ile Glu Arg Val Pro Tyr Ala Lys Pro	217
15 20 25	
CCT TTC ACC ATC AGC GAC CTC AAA AAA GCC ATT CCT CCC CAC TGT TTC Pro Phe Thr Ile Ser Asp Leu Lys Ala Ile Pro Pro His Cys Phe	265
30 35 40	
CAG CGT TCC CTT ATC CGT TCC TTC TCT TAT GTC GTT TAT GAC CTT GCT Gln Arg Ser Leu Ile Arg Ser Phe Ser Tyr Val Val Tyr Asp Leu Ala	313
45 50 55 60	
GTG AGC TTC CTC CTC TAC TAT GTA GCC GCC ACT TAC TTC CAC CAT CTG Val Ser Phe Leu Leu Tyr Tyr Val Ala Ala Thr Tyr Phe His His Leu	361
65 70 75	
CCA AAC CCT TTC TCC CTT GCG TGG CTG GCT TAT TGG GTC GTT CAA Pro Asn Pro Phe Ser Ser Leu Ala Trp Leu Ala Tyr Trp Val Val Gln	409
80 85 90	
GGC TGT GTG CTT ACA GGA GTG TGG GTC ATA GCC CAT GAA TGT GGT CAC Gly Cys Val Leu Thr Gly Val Trp Val Ile Ala His Glu Cys Gly His	457
95 100 105	
CAT GCA TTT AGT GAC TAT CAA TGG GTT GAT GAC ACT GTG GGC TTC CTA His Ala Phe Ser Asp Tyr Gln Trp Val Asp Asp Thr Val Gly Phe Leu	505
110 115 120	
CTC CAC TCG GTT CTA CTT GTT CCT TTC TTT TCA TGG AAA TAC AGT CAT Leu His Ser Val Leu Val Pro Phe Phe Ser Trp Lys Tyr Ser His	553
125 130 135 140	
CGT CGA CAC CAC TCC AAC ACC GGA TCA CTC GAG CGT GAT GAA GTC TTT Arg Arg His His Ser Asn Thr Gly Ser Leu Glu Arg Asp Glu Val Phe	601
145 150 155	
GTC CCA AAA CCG AGA TCG AAA ATC CCT TGG TAC TCA AAA TAC TTT AAC Val Pro Lys Pro Arg Ser Lys Ile Pro Trp Tyr Ser Lys Tyr Phe Asn	649
160 165 170	

AAC GCA CCT GGC CGC ATG ATG AGT GTG TTC ACC ACC CTA ACT CTA GGC Asn Ala Pro Gly Arg Met Met Ser Val Phe Thr Thr Leu Thr Leu Gly 175 180 185	697
TGG CCC TTG TAC TTG GTT TTC AAT GTA TCA GGG AGA CCC TAT GAC CGT Trp Pro Leu Tyr Leu Val Phe Asn Val Ser Gly Arg Pro Tyr Asp Arg 190 195 200	745
TTT GCC TGC CAC TTT TCT CCT AAC AGC CCT ATA TAC AAC GAA CGT GAG Phe Ala Cys His Phe Ser Pro Asn Ser Pro Ile Tyr Asn Glu Arg Glu 205 210 215 220	793
CGT CTC CAA ATA TGG CTT TCG GAT TTA GGG ATG ATC ACC ATG TCG TTC Arg Leu Gln Ile Trp Leu Ser Asp Leu Gly Met Ile Thr Met Ser Phe 225 230 235	841
ATC CTT TAT CGT GTT GCT GTA GCA AAA GGT GTG GCT TGG GTA ATA TGC Ile Leu Tyr Arg Val Ala Val Ala Lys Gly Val Ala Trp Val Ile Cys 240 245 250	889
ATG TAT GGG ATC CCG CTA CTG ATT GTG AAC GGA TTC CTG GTG ACG ATC Met Tyr Gly Ile Pro Leu Leu Ile Val Asn Gly Phe Leu Val Thr Ile 255 260 265	937
ACG TAC CTT CAA CAC ACT CAC CCT TCA TTG CCC CAC TAT GAT AGC TCA Thr Tyr Leu Gln His Thr His Pro Ser Leu Pro His Tyr Asp Ser Ser 270 275 280	985
GAG TGG GAC TGG CTA AGG GGA GCA ATG GCA ACG GTG GAC CGT GAC TAT Glu Trp Asp Trp Leu Arg Gly Ala Met Ala Thr Val Asp Arg Asp Tyr 285 290 295 300	1033
GGT GTG CTC AAC AAG GTA TTC CAT AAC ATC ACA GAT ACA CAC GTG GTG Gly Val Leu Asn Lys Val Phe His Asn Ile Thr Asp Thr His Val Val 305 310 315	1081
CAC CAT TTG TTC TCG ACG ATG CCT CAT TAT AAC GCA ATG GAG GCA ACG His His Leu Phe Ser Thr Met Pro His Tyr Asn Ala Met Glu Ala Thr 320 325 330	1129
AAA GCA GTG AAG CCT TTG CTT GGG GAG TAT TAT CAG TTT GAC GGG ACT Lys Ala Val Lys Pro Leu Leu Gly Glu Tyr Tyr Gln Phe Asp Gly Thr 335 340 345	1177
CCG TTT TAC GTA GCA ATA TGG AGA GAG GCA AAG GAG TGT CTG TTC GTG Pro Phe Tyr Val Ala Ile Trp Arg Glu Ala Lys Glu Cys Leu Phe Val 350 355 360	1225
GAT CCA GAT GAG GGG GAG GGT CAG GGA GGT GTG TTT TGG TAC AAG AAT Asp Pro Asp Glu Gly Glu Gly Gln Gly Val Phe Trp Tyr Lys Asn 365 370 375 380	1273
AAG ATG TGATATTCA ATGATGAACA AGTTTATAG GTTATCGATC AGATCAGGTC Lys Met	1329
GGTTTATGTT GTTTATGTGC TTATGTGTGT GTGTGTTTT TATGTGTAGC AAGCAGTTGA	1389
TCGAGTGTG GGTATGTATG TGTCGTAGCG TCAATGAATT GAAAAGAACT GGTTATGTTC	1449
TTTAACATCA AAAAAAAA AAAAAAA	1476

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 382 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Gly Ala Gly Gly Cys Met Ser Ala Ser Glu Thr Lys Thr Gln Gln
 1 5 10 15

Lys Asn Pro Ile Glu Arg Val Pro Tyr Ala Lys Pro Pro Phe Thr Ile
 20 25 30

Ser Asp Leu Lys Lys Ala Ile Pro Pro His Cys Phe Gln Arg Ser Leu
 35 40 45

Ile Arg Ser Phe Ser Tyr Val Val Tyr Asp Leu Ala Val Ser Phe Leu
 50 55 60

Leu Tyr Tyr Val Ala Ala Thr Tyr Phe His His Leu Pro Asn Pro Phe
 65 70 75 80

Ser Ser Leu Ala Trp Leu Ala Tyr Trp Val Val Gln Gly Cys Val Leu
 85 90 95

Thr Gly Val Trp Val Ile Ala His Glu Cys Gly His His Ala Phe Ser
 100 105 110

Asp Tyr Gln Trp Val Asp Asp Thr Val Gly Phe Leu Leu His Ser Val
 115 120 125

Leu Leu Val Pro Phe Phe Ser Trp Lys Tyr Ser His Arg Arg His His
 130 135 140

Ser Asn Thr Gly Ser Leu Glu Arg Asp Glu Val Phe Val Pro Lys Pro
 145 150 155 160

Arg Ser Lys Ile Pro Trp Tyr Ser Lys Tyr Phe Asn Asn Ala Pro Gly
 165 170 175

Arg Met Met Ser Val Phe Thr Thr Leu Thr Leu Gly Trp Pro Leu Tyr
 180 185 190

Leu Val Phe Asn Val Ser Gly Arg Pro Tyr Asp Arg Phe Ala Cys His
 195 200 205

Phe Ser Pro Asn Ser Pro Ile Tyr Asn Glu Arg Glu Arg Leu Gln Ile
 210 215 220

Trp Leu Ser Asp Leu Gly Met Ile Thr Met Ser Phe Ile Leu Tyr Arg
 225 230 235 240

Val Ala Val Ala Lys Gly Val Ala Trp Val Ile Cys Met Tyr Gly Ile
 245 250 255

Pro Leu Leu Ile Val Asn Gly Phe Leu Val Thr Ile Thr Tyr Leu Gln
 260 265 270

His Thr His Pro Ser Leu Pro His Tyr Asp Ser Ser Glu Trp Asp Trp
 275 280 285

Leu Arg Gly Ala Met Ala Thr Val Asp Arg Asp Tyr Gly Val Leu Asn
 290 295 300

Lys Val Phe His Asn Ile Thr Asp Thr His Val Val His His Leu Phe
 305 310 315 320
 Ser Thr Met Pro His Tyr Asn Ala Met Glu Ala Thr Lys Ala Val Lys
 325 330 335
 Pro Leu Leu Gly Glu Tyr Tyr Gln Phe Asp Gly Thr Pro Phe Tyr Val
 340 345 350
 Ala Ile Trp Arg Glu Ala Lys Glu Cys Leu Phe Val Asp Pro Asp Glu
 355 360 365
 Gly Glu Gly Gln Gly Val Phe Trp Tyr Lys Asn Lys Met
 370 375 380

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1364 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 103..1254
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

TTATGAAAGC TCGATCGGTG TTTCGATCAAT TCAAATCGAC GAACACGAAA TCGAACTCAA	60
CAATTCAAAT CTGGAAATAT TAATTGGATC AAGCGGGCGG AT ATG ATG ATG TCG Met Met Met Ser	114
1	
GAT TCA TGT GAT GAT CAT GAT CAG CTG GTG AAA GAT GAT CAT AAT ATA Asp Ser Cys Asp Asp His Asp Gln Leu Val Lys Asp Asp His Asn Ile	162
5 10 15 20	
AAC GAA CGT GCA CCG GTT GAT GCG GCA CCA TTC TCG TTA AGC GAT CTA Asn Glu Arg Ala Pro Val Asp Ala Ala Pro Phe Ser Leu Ser Asp Leu	210
25 30 35	
AAG AAA GCA ATC CCT CCG CAT TGC TTC CAG CGA TCT GCC ATC CGT TCA Lys Lys Ala Ile Pro Pro His Cys Phe Gln Arg Ser Ala Ile Arg Ser	258
40 45 50	
TCG TGC TAC GTT CAG GAT CTC ATT ATT ACC TTC CTT TTA TAC ACG Ser Cys Tyr Val Val Gln Asp Leu Ile Ile Thr Phe Leu Leu Tyr Thr	306
55 60 65	
CTC GCC AAC TCT TAC ATT CCT CTT CTT CCT CCT CTA CCT TAC TTA Leu Ala Asn Ser Tyr Ile Pro Leu Leu Pro Pro Leu Pro Tyr Leu	354
70 75 80	
GCA TGG CCT GTT TAC TGG TTT TGC CAA TCT TCG ATC CTC ACT GGT TTA Ala Trp Pro Val Tyr Trp Phe Cys Gln Ser Ser Ile Leu Thr Gly Leu	402
85 90 95 100	
TGG GTC ATT GGC CAT GAA TGT GGC CAT CAT GCT TAT AGT GAG TAC CAG Trp Val Ile Gly His Glu Cys Gly His Ala Tyr Ser Glu Tyr Gln	450
105 110 115	

TGG GTT GAT AAC ACC GTT GGA TTC ATC CTC CAT TCC TTT CTT CTC ACA Trp Val Asp Asn Thr Val Gly Phe Ile Leu His Ser Phe Leu Leu Thr 120 125 130	498
CCT TAC TTT TCT TGG AAA TAC AGC CAT CGA AAG CAC CAT GCC AAC ACG Pro Tyr Phe Ser Trp Lys Tyr Ser His Arg Lys His His Ala Asn Thr 135 140 145	546
AAT TCA CTC GAA AAC GAG GAG GTT TAC ATT CCT AAA GCC AAG TCC CAG Asn Ser Leu Glu Asn Glu Glu Val Tyr Ile Pro Lys Ala Lys Ser Gln 150 155 160	594
CTC AGG AAT TAC TCC AAT TTC AAA TTT CTT GAC AAC ACC CCT GGT CGA Leu Arg Asn Tyr Ser Asn Phe Lys Phe Leu Asp Asn Thr Pro Gly Arg 165 170 175 180	642
ATC TTC ATT TTG CTT ATC ATG TTG ACC TTG GGC TTT CCT TTA TAC CTC Ile Phe Ile Leu Ile Met Leu Thr Leu Gly Phe Pro Leu Tyr Leu 185 190 195	690
TTG ACC AAT ATT TCA GGC AAG AAA TAC CAA AGG TTT GCC AAC CAC TTT Leu Thr Asn Ile Ser Gly Lys Lys Tyr Gln Arg Phe Ala Asn His Phe 200 205 210	738
GAT CCG TTG AGC CCC ATC TTC AGT GAG CGT GAA CGA ATC CAG GTC GTG Asp Pro Leu Ser Pro Ile Phe Ser Glu Arg Glu Arg Ile Gln Val Val 215 220 225	786
CTA TCG GAT GTG GGT CTC ATT GCT GTG TTT TAC GGG CTT AAG TTT CTT Leu Ser Asp Val Gly Leu Ile Ala Val Phe Tyr Gly Leu Lys Phe Leu 230 235 240	834
GTA GCG AAA AAA GGG TTC GGT TGG GTA ATG CGC ATG TAC GGA GCC CCA Val Ala Lys Lys Gly Phe Gly Trp Val Met Arg Met Tyr Gly Ala Pro 245 250 255 260	882
GTG GTT GGG CTG AAT GCC TTC ATA ATA ATG ATC ACT TAT CTC CAC CAC Val Val Gly Leu Asn Ala Phe Ile Ile Met Ile Thr Tyr Leu His His 265 270 275	930
ACC CAT CTG TCT TCG CCT CAT TAC GAT TCG ACC GAA TGG AAC TGG ATC Thr His Leu Ser Ser Pro His Tyr Asp Ser Thr Glu Trp Asn Trp Ile 280 285 290	978
AAA GGA GCC TTG ACT ACA ATC GAT AGA GAT TTC GGT CTC CTG AAT AGG Lys Gly Ala Leu Thr Thr Ile Asp Arg Asp Phe Gly Leu Leu Asn Arg 295 300 305	1026
GTG TTC CAT GAC GTC ACT CAC ACA CAC GTG TTG CAT CAT TTG TTC CCG Val Phe His Asp Val Thr His Thr His Val Leu His His Leu Phe Pro 310 315 320	1074
TAC ATT CCA CAT TAT CAT GCA AAG GAG GCG AGC GAC GCA ATA AAG CCG Tyr Ile Pro His Tyr His Ala Lys Glu Ala Ser Asp Ala Ile Lys Pro 325 330 335 340	1122
GTG TTA GGG GAG TAT CGG ATG ATC GAT AGG ACT CCG TTT TAC AAA GCA Val Leu Gly Glu Tyr Arg Met Ile Asp Arg Thr Pro Phe Tyr Lys Ala 345 350 355	1170
ATG TGG AGA GAG GCG AAG GAA TGC ATC TAC ATC GAG CCA GAT GAA GAT Met Trp Arg Glu Ala Lys Glu Cys Ile Tyr Ile Glu Pro Asp Glu Asp 360 365 370	1218

AAG AAG CAC AAA GGT GTA TAT TGG TAC CAT AAA ATG TGATACGAGC Lys Lys His Lys Gly Val Tyr Trp Tyr His Lys Met 375 380	1264
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TGAGTACGTA GTACGTTGTA TGCTTTGTA ACGTTTGTA AGATAAATAA ATAAATCTTG	1324
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AATGAAGATA AAAAAAAA AAAAAAAA AAAAAAAA	1364
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(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 384 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Met Met Ser Asp Ser Cys Asp Asp His Asp Gln Leu Val Lys Asp 1 5 10 15
--

Asp His Asn Ile Asn Glu Arg Ala Pro Val Asp Ala Ala Pro Phe Ser 20 25 30

Leu Ser Asp Leu Lys Lys Ala Ile Pro Pro His Cys Phe Gln Arg Ser 35 40 45

Ala Ile Arg Ser Ser Cys Tyr Val Val Gln Asp Leu Ile Ile Thr Phe 50 55 60

Leu Leu Tyr Thr Leu Ala Asn Ser Tyr Ile Pro Leu Leu Pro Pro Pro 65 70 75 80
--

Leu Pro Tyr Leu Ala Trp Pro Val Tyr Trp Phe Cys Gln Ser Ser Ile 85 90 95

Leu Thr Gly Leu Trp Val Ile Gly His Glu Cys Gly His His Ala Tyr 100 105 110
--

Ser Glu Tyr Gln Trp Val Asp Asn Thr Val Gly Phe Ile Leu His Ser 115 120 125
--

Phe Leu Leu Thr Pro Tyr Phe Ser Trp Lys Tyr Ser His Arg Lys His 130 135 140
--

His Ala Asn Thr Asn Ser Leu Glu Asn Glu Glu Val Tyr Ile Pro Lys 145 150 155 160
--

Ala Lys Ser Gln Leu Arg Asn Tyr Ser Asn Phe Lys Phe Leu Asp Asn 165 170 175
--

Thr Pro Gly Arg Ile Phe Ile Leu Ile Met Leu Thr Leu Gly Phe 180 185 190
--

Pro Leu Tyr Leu Leu Thr Asn Ile Ser Gly Lys Lys Tyr Gln Arg Phe 195 200 205
--

Ala Asn His Phe Asp Pro Leu Ser Pro Ile Phe Ser Glu Arg Glu Arg 210 215 220
--

Ile Gln Val Val Leu Ser Asp Val Gly Leu Ile Ala Val Phe Tyr Gly 225 230 235 240
--

Leu Lys Phe Leu Val Ala Lys Lys Gly Phe Gly Trp Val Met Arg Met 245 250 255
--

Tyr Gly Ala Pro Val Val Gly Leu Asn Ala Phe Ile Ile Met Ile Thr
 260 265 270
 Tyr Leu His His Thr His Leu Ser Ser Pro His Tyr Asp Ser Thr Glu
 275 280 285
 Trp Asn Trp Ile Lys Gly Ala Leu Thr Thr Ile Asp Arg Asp Phe Gly
 290 295 300
 Leu Leu Asn Arg Val Phe His Asp Val Thr His Thr His Val Leu His
 305 310 315 320
 His Leu Phe Pro Tyr Ile Pro His Tyr His Ala Lys Glu Ala Ser Asp
 325 330 335
 Ala Ile Lys Pro Val Leu Gly Glu Tyr Arg Met Ile Asp Arg Thr Pro
 340 345 350
 Phe Tyr Lys Ala Met Trp Arg Glu Ala Lys Glu Cys Ile Tyr Ile Glu
 355 360 365
 Pro Asp Glu Asp Lys Lys His Lys Gly Val Tyr Trp Tyr His Lys Met
 370 375 380

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 387 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met Gly Leu Ala Lys Glu Thr Thr Met Gly Gly Arg Gly Arg Val Ala
 1 5 10 15
 Lys Val Glu Val Gln Gly Lys Lys Pro Leu Ser Arg Val Pro Asn Thr
 20 25 30
 Lys Pro Pro Phe Thr Val Gly Gln Leu Lys Lys Ala Ile Pro Pro His
 35 40 45
 Cys Phe Gln Arg Ser Leu Leu Thr Ser Phe Ser Tyr Val Val Tyr Asp
 50 55 60
 Leu Ser Phe Ala Phe Ile Phe Tyr Ile Ala Thr Thr Tyr Phe His Leu
 65 70 75 80
 Leu Pro Gln Pro Phe Ser Leu Ile Ala Trp Pro Ile Tyr Trp Val Leu
 85 90 95
 Gln Gly Cys Leu Leu Thr Gly Val Trp Val Ile Ala His Glu Cys Gly
 100 105 110
 His His Ala Phe Ser Lys Tyr Gln Trp Val Asp Asp Val Val Gly Leu
 115 120 125
 Thr Leu His Ser Thr Leu Leu Val Pro Tyr Phe Ser Trp Lys Ile Ser
 130 135 140
 His Arg Arg His His Ser Asn Thr Gly Ser Leu Asp Arg Asp Glu Val
 145 150 155 160

Phe Val Pro Lys Pro Lys Ser Lys Val Ala Trp Phe Ser Lys Tyr Leu
 165 170 175
 Asn Asn Pro Leu Gly Arg Ala Val Ser Leu Leu Val Thr Leu Thr Ile
 180 185 190
 Gly Trp Pro Met Tyr Leu Ala Phe Asn Val Ser Gly Arg Pro Tyr Asp
 195 200 205
 Ser Phe Ala Ser His Tyr His Pro Tyr Ala Pro Ile Tyr Ser Asn Arg
 210 215 220
 Glu Arg Leu Leu Ile Tyr Val Ser Asp Val Ala Leu Phe Ser Val Thr
 225 230 235 240
 Tyr Ser Leu Tyr Arg Val Ala Thr Leu Lys Gly Leu Val Trp Leu Leu
 245 250 255
 Cys Val Tyr Gly Val Pro Leu Leu Ile Val Asn Gly Phe Leu Val Thr
 260 265 270
 Ile Thr Tyr Leu Gln His Thr His Phe Ala Leu Pro His Tyr Asp Ser
 275 280 285
 Ser Glu Trp Asp Trp Leu Lys Gly Ala Leu Ala Thr Met Asp Arg Asp
 290 295 300
 Tyr Gly Ile Leu Asn Lys Val Phe His His Ile Thr Asp Thr His Val
 305 310 315 320
 Ala His His Leu Phe Ser Thr Met Pro His Tyr His Ala Met Glu Ala
 325 330 335
 Thr Asn Ala Ile Lys Pro Ile Leu Gly Glu Tyr Tyr Gln Phe Asp Asp
 340 345 350
 Thr Pro Phe Tyr Lys Ala Leu Trp Arg Glu Ala Arg Glu Cys Leu Tyr
 355 360 365
 Val Glu Pro Asp Glu Gly Thr Ser Glu Lys Gly Val Tyr Trp Tyr Arg
 370 375 380
 Asn Lys Tyr
 385

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 387 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Gly Gly Gly Gly Arg Met Ser Thr Val Ile Thr Ser Asn Asn Ser
 1 5 10 15
 Glu Lys Lys Gly Gly Ser Ser His Leu Lys Arg Ala Pro His Thr Lys
 20 25 30
 Pro Pro Phe Thr Leu Gly Asp Leu Lys Arg Ala Ile Pro Pro His Cys
 35 40 45

Phe Glu Arg Ser Phe Val Arg Ser Phe Ser Tyr Val Ala Tyr Asp Val
 50 55 60
 Cys Leu Ser Phe Leu Phe Tyr Ser Ile Ala Thr Asn Phe Phe Pro Tyr
 65 70 75 80
 Ile Ser Ser Pro Leu Ser Tyr Val Ala Trp Leu Val Tyr Trp Leu Phe
 85 90 95
 Gln Gly Cys Ile Leu Thr Gly Leu Trp Val Ile Gly His Glu Cys Gly
 100 105 110
 His His Ala Phe Ser Glu Tyr Gln Leu Ala Asp Asp Ile Val Gly Leu
 115 120 125
 Ile Val His Ser Ala Leu Leu Val Pro Tyr Phe Ser Trp Lys Tyr Ser
 130 135 140
 His Arg Arg His His Ser Asn Ile Gly Ser Leu Glu Arg Asp Glu Val
 145 150 155 160
 Phe Val Pro Lys Ser Lys Ser Lys Ile Ser Trp Tyr Ser Lys Tyr Ser
 165 170 175
 Asn Asn Pro Pro Gly Arg Val Leu Thr Leu Ala Ala Thr Leu Leu Leu
 180 185 190
 Gly Trp Pro Leu Tyr Leu Ala Phe Asn Val Ser Gly Arg Pro Tyr Asp
 195 200 205
 Arg Phe Ala Cys His Tyr Asp Pro Tyr Gly Pro Ile Phe Ser Glu Arg
 210 215 220
 Glu Arg Leu Gln Ile Tyr Ile Ala Asp Leu Gly Ile Phe Ala Thr Thr
 225 230 235 240
 Phe Val Leu Tyr Gln Ala Thr Met Ala Lys Gly Leu Ala Trp Val Met
 245 250 255
 Arg Ile Tyr Gly Val Pro Leu Leu Ile Val Asn Cys Phe Leu Val Met
 260 265 270
 Ile Thr Tyr Leu Gln His Thr His Pro Ala Ile Pro Arg Tyr Gly Ser
 275 280 285
 Ser Glu Trp Asp Trp Leu Arg Gly Ala Met Val Thr Val Asp Arg Asp
 290 295 300
 Tyr Gly Val Leu Asn Lys Val Phe His Asn Ile Ala Asp Thr His Val
 305 310 315 320
 Ala His His Leu Phe Ala Thr Val Pro His Tyr His Ala Met Glu Ala
 325 330 335
 Thr Lys Ala Ile Lys Pro Ile Met Gly Glu Tyr Tyr Arg Tyr Asp Gly
 340 345 350
 Thr Pro Phe Tyr Lys Ala Leu Trp Arg Glu Ala Lys Glu Cys Leu Phe
 355 360 365
 Val Glu Pro Asp Glu Gly Ala Pro Thr Gln Gly Val Phe Trp Tyr Arg
 370 375 380
 Asn Lys Tyr
 385

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 30 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

ATGCGGCCGC ATGATGATGT CGGATTCTATG

30

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 32 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

ATGCGGCCGC TCACATTTA TGGTACCAAT AT

32

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 26 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

ATGCGGCCGC CATGGGAGCA GGAGGG

26

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 32 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

ATGCGGCCGC TCACATCTTA TTCTTGATCC AA

32

INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>5</u> , line <u>6</u>	
B. IDENTIFICATION OF DEPOSIT	
Name of depositary institution AMERICAN TYPE CULTURE COLLECTION	
Address of depositary institution (including postal code and country) 10801 University Blvd. Manassas, Virginia 20110-2209 USA	
Date of deposit	Accession Number
C. ADDITIONAL INDICATIONS (leave blank if not applicable)	
In respect of those designations in which a European patent is sought, a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample. (Rule 28(4) EPC)	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit") Accession number and number of deposit will be submitted later	

For receiving Office use only	
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Authorized officer	

CLAIMS

What is claimed is:

1. An isolated nucleic acid fragment encoding a *Vernonia galamenensis* fatty acid desaturase comprising a member selected from the group consisting of:
 - 5 (a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:2;
 - (b) an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:2; and
 - 10 (c) an isolated nucleic acid fragment that is complementary to (a) or (b).
2. The isolated nucleic acid fragment of Claim 1 wherein the nucleotide sequence of the fragment is set forth in SEQ ID NO:1.
3. A chimeric gene comprising the nucleic acid fragment of Claim 1 operably linked to suitable regulatory sequences.
- 15 4. A transformed host cell comprising the chimeric gene of Claim 3.
5. An isolated nucleic acid fragment encoding a *Vernonia galamenensis* fatty acid epoxidizing enzyme comprising a member selected from the group consisting of:
 - (a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:4;
 - 20 (b) an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:4; and
 - (c) an isolated nucleic acid fragment that is complementary to (a) or (b).
6. The isolated nucleic acid fragment of Claim 5 wherein the nucleotide sequence 25 of the fragment is set forth in SEQ ID NO:3.
7. A chimeric gene comprising the nucleic acid fragment of Claim 5 operably linked to suitable regulatory sequences.
8. A transformed host cell comprising the chimeric gene of Claim 7.
9. A transformed plant comprising the host cell of Claim 8.
- 30 10. A seed from the transformed plant of Claim 9 wherein the seed comprises the chimeric gene of Claim 7.
11. A soybean seed that accumulates vernolic acid.
12. The soybean seed of Claim 11 wherein said seed bears ATCC Accession No. _____.
- 35 13. A method of altering the level of expression of a *Vernonia galamenensis* fatty acid modifying enzyme in a host cell comprising:
 - (a) transforming a host cell with the chimeric gene of Claim 3 or the chimeric gene of Claim 7; and

(b) growing the transformed host cell produced in step (a) under conditions that are suitable for expression of the chimeric gene

wherein expression of the chimeric gene results in production of altered levels of a *Vernonia galamenensis* fatty acid modifying enzyme in the transformed host cell.

5 14. A method for producing epoxidized fatty acids in the seeds of plants comprising the following steps:

(a) transforming a plant cell with a chimeric gene encoding all or a portion of a *Vernonia galamenensis* fatty acid epoxidizing enzyme operably linked in sense orientation to suitable regulatory sequences;

10 (b) growing a fertile mature plant from the transformed plant cell of step (a) under conditions suitable to obtain seeds; and

(c) selecting from the progeny seed of step (b) those seeds containing epoxidized fatty acids.

15 15. A method for producing a *Vernonia galamenensis* fatty acid modifying enzyme comprising the following steps:

(a) transforming a microbial host cell with a chimeric gene wherein a nucleic acid fragment encoding a *Vernonia galamenensis* fatty acid modifying enzyme is operably linked to regulatory sequences suitable for directing expression in microbial cells; and

20 (b) growing the transformed microbial cells obtained from step (a) under conditions that result in expression of the *Vernonia galamenensis* fatty acid modifying enzyme protein.

25 16. A method of obtaining a nucleic acid fragment encoding all or substantially all of the amino acid sequence encoding a *Vernonia galamenensis* fatty acid modifying enzyme comprising:

(a) probing a cDNA or genomic library with the nucleic acid fragment of Claim 1 or Claim 5;

(b) identifying a DNA clone that hybridizes to the nucleic acid fragment of Claim 1 or Claim 5; and

30 (c) isolating the DNA clone identified in step (b);

wherein the isolated DNA clone of step (c) comprises a nucleic acid fragment encoding all or substantially all of the amino acid sequence encoding a *Vernonia galamenensis* fatty acid modifying enzyme.

35 17. A method of obtaining a nucleic acid fragment encoding a portion of an amino acid sequence encoding a *Vernonia galamenensis* fatty acid modifying enzyme comprising:

(a) synthesizing an oligonucleotide primer corresponding to a portion of the sequence set forth in a member selected from the group consisting of SEQ ID NO:1 and SEQ ID NO:3; and

(b) amplifying a cDNA insert present in a cloning vector using the oligonucleotide primer of step (a) and a primer representing sequences of the cloning vector

wherein the amplified nucleic acid fragment encodes a portion of an amino acid sequence
5 encoding a *Vernonia galamenensis* fatty acid modifying enzyme.

18. The product of the method of Claim 16.

19. The product of the method of Claim 17.

1 MGLAKETTMGGRGRVAKVEVOGKKPLSRVPNTKPPFTVGQLKKAIAPPHCFORSLILTSFSY SOY
 1 ----- MGAGGCMSETKTKQQKNPPIERVPYAKPPFTISDLKKAIAPPHCFORSLIRFSY vs1.05h08
 1 ----- MMSDSCDDHDQLVKDDHINERAPVDAAPFSLSLDLKKAIAPPHCFORSAIRSSCY vs1.02c07
 1 -MGGGRMSTVITSNNSEKKGGSSHLKRAPHTKPPFTLGLKRAAPPHCERSVRSFSY castor

61 VVYDLSFAFI - FYIATTYFILLPOPPESLIAWPIYVWVQGLLIGVWVIA [REMOVED] HAFSKY SOY
 55 VVYDLAWSFLYVVAATYFHHLPNPFSLLVPIPLPPPIKLAWPVYWFQSSLILTGLWV [REMOVED] HAFSDY vs1.05h08
 56 VVQDLIITFLYTLANSYIPLLPPPIKLAWPVYWFQSSLILTGLWV [REMOVED] HAYSEY vs1.02c07
 60 VAYDVCLSLFLFYSIATNFFPYISSLSPLSYVAWLVYWLFOGCCILTGLWV [REMOVED] HAFSEY castor

120 QWVDDVVGLTLHSTLLVPPYFSWKIS [REMOVED] SNTGSLDRDEVFVVKPKSKVVAWFS--KYLN SOY
 115 QWVDDDTVGFLHSVLLVPPFSWKYS [REMOVED] SNTGSLERDEVFVVKPRSKIPWYS--KYFN vs1.05h08
 116 QWVTN TVGFLHSFLLTPYFSWKYS [REMOVED] ANTNSILENEEVYIPKAKS [REMOVED] LRNYSNFKF [REMOVED] vs1.02c07
 120 QLADDIVGLIVHSALLVPPYFSWKYS [REMOVED] SNIGSLERDEVFVVKSKSKISWYS--KYSN castor

178 NPLGRAVSLLIVTLLTIGWPMYLAFLNVSGRPyDSFASHYHYPYAPIYSNRERLLIYVSDVALF SOY
 173 NAPGRMMSVFTTLGWPPLYLVFTNVSGRPyDRFACHFSPNSPIYNERERLQIWLSDLGMI vs1.05h08
 176 NTPGRIFILLIMLTLGFPLYLNTNISGM [REMOVED] YQRFANHFDPLSPIFI SERERIQLVVLSDVGLI vs1.02c07
 178 NPPGRVLTAAATLLGWPPLYLAFLNVSGRPyDRFACHYDPYGPIFI SERERLQIYIADLGIF castor

238 SVTYSLYRVATLKGVLWILLCVYGVPLLIVINGFLVTITYLQHOTHFALPHYDSSEWDWLKGKA SOY
 233 TMSFLILYRVAVAKGVAAWVICMYGIPLLIVNGFLVTITYLQHOTHPSLPHYDSSEWDWLRLGA vs1.05h08
 236 AVFYGLKFLVAKKGFGWV [REMOVED] MYGAPVVGLNANFLIMITY [REMOVED] RTHLSSPHYDSTEM [REMOVED] WIKGA vs1.02c07
 238 ATTFLVYQATMAKGLAWV [REMOVED] IYGVPLLIVNCFLVMTYLQHOTHPAIPRGSSSEWDWLRLGA castor

298 LATMDRDYGILNKVFHHITD [REMOVED] LFSTTMPHYHAMEATNAIKPILGEYYQFDDTPFYK SOY
 293 MATVDRDYGVLNKVFHN [REMOVED] LFSTTMPHYHAMEATKAVKPLLGEYYQFDGTPFYV vs1.05h08
 296 LTTIDRDFGLLNRFVHDVTH [REMOVED] LFPTYIIPHYHAMEASDAIKPVLGEYRMDRTPFYK vs1.02c07
 298 MVTVDRDYGVLNKVFHNIA [REMOVED] LFATVPHYHAMEATKAIAKPIIMEYYRYDGTGTPFYK castor

358 ALWREARECLYVEPDEGTSEKGVWYRNKY SOY
 353 AIWREAKECLFVDPDEGEQGGGVFWYKNKM vs1.05h08
 356 AMWREAKECLFVDPDEGEKHHKGVYMYH - KM vs1.02c07
 358 ALWREAKECLFVEPDEGAPTOGVFWYRNK - Y castor

FIGURE 2

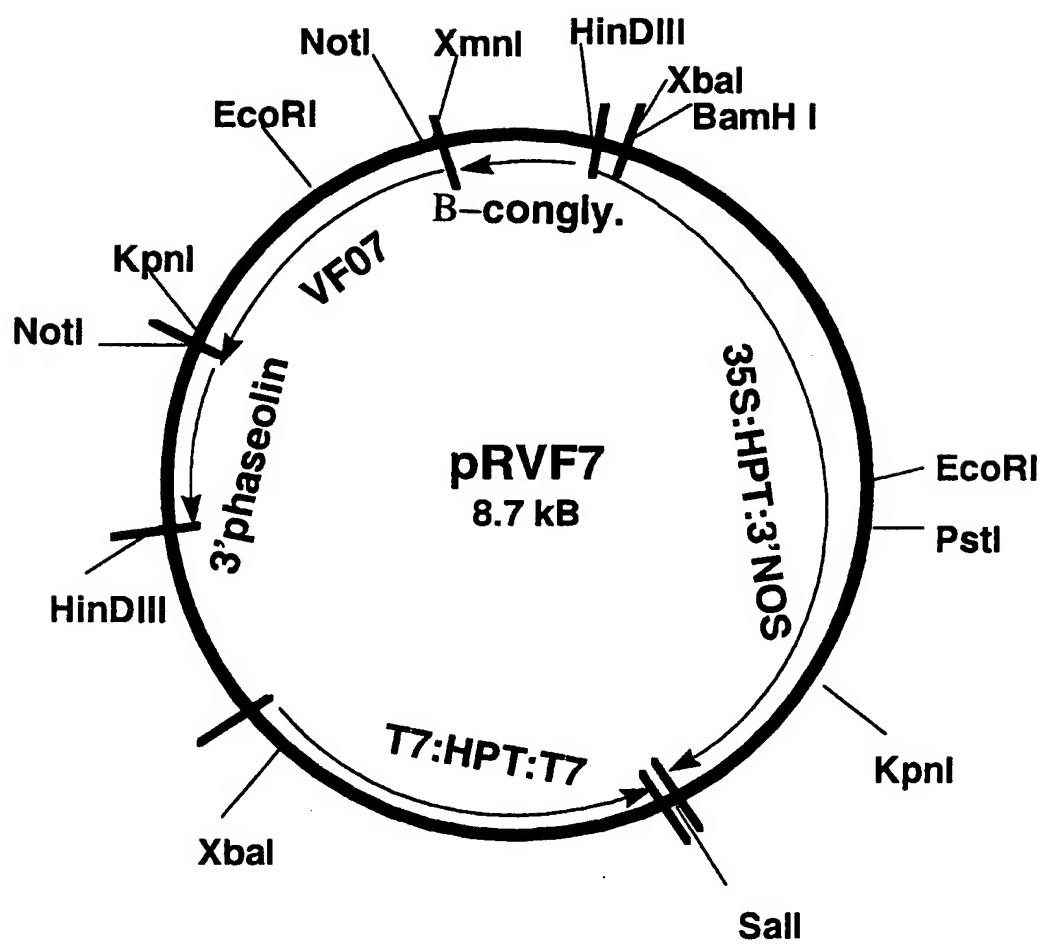
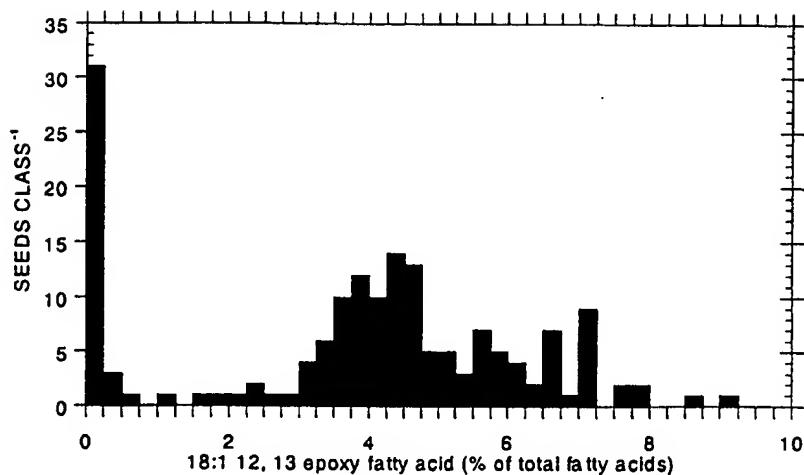
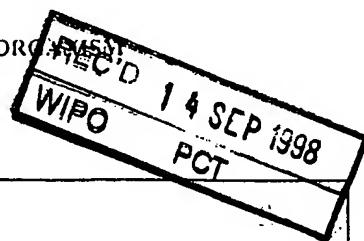


FIGURE 3

INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)



A. The indications made below relate to the microorganism referred to in the description
on page 5, line 6

B. IDENTIFICATION OF DEPOSIT

Further deposits are identified on an additional sheet

Name of depository institution

AMERICAN TYPE CULTURE COLLECTION

Address of depository institution (including postal code and country)

10801 University Blvd.
Manassas, Virginia 20110-2209
USA

Date of deposit

13 June 1998 (13.06.98)

Accession Number

209967

C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet

In respect of those designations in which a European patent is sought, a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample. (Rule 28(4) EPO)

D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)

E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)

The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g. "Accession Number of Deposit")

Accession and Number of Deposit will be submitted later

For receiving Office use only

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For International Bureau use only

 This sheet was received by the International Bureau on:

14 SEP 1998

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Authorized officer

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 98/12074

A. CLASSIFICATION OF SUBJECT MATTER					
IPC 6	C12N15/53	C12N15/11	C12N15/82	C12N1/21	C12N5/10
	C12N9/02	C12Q1/68	A01H5/00		

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C12Q A01H

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category ^a	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	SEITHER G. ET AL.: "Isolation of cytochrome P-450 genes from Vernonia galamensis" PHYSIOL. BIOCHEM. MOL. BIOL. PLANT LIPIDS (PROC. INT. SYMP. PLANT LIPIDS), 1997, pages 389-391, XP002080169 see the whole document ---	5, 7-10, 15, 16
A	BAFOR M. ET AL.: "Biosynthesis of vernoleate (cis-12-epoxyoctadeca-cis-9-enoate) in microsomal preparations from developing endosperm of Euphorbia lagascae" ARCHIVES OF BIOCHEMISTRY AND BIOPHYSICS, vol. 303, no. 1, 15 May 1993, pages 145-151, XP002080170 cited in the application see the whole document ---	1-19

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

^a Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

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Date of the actual completion of the international search

9 October 1998

Date of mailing of the international search report

22/10/1998

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Kania, T

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 98/12074

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 94 11516 A (DU PONT ;LIGHTNER JONATHAN EDWARD (US); OKULEY JOHN JOSEPH (US)) 26 May 1994 cited in the application see the whole document -----	1-19
A	US 5 430 134 A (OHLROGGE JOHN B ET AL) 4 July 1995 see the whole document -----	1-19

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 98/12074

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
WO 9411516	A 26-05-1994	AU 5407594	A 08-06-1994	
		CA 2149223	A 26-05-1994	
		EP 0668919	A 30-08-1995	
		JP 8503364	T 16-04-1996	
US 5430134	A 04-07-1995	NONE		